

Update on environmental DNA (eDNA) monitoring of *Aphanomyces astaci* and freshwater crayfish



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Head of Fish Health Research Group

10th AW of the NRL for Crustacean Diseases – May 29th 2019



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The Fish Health Research Group



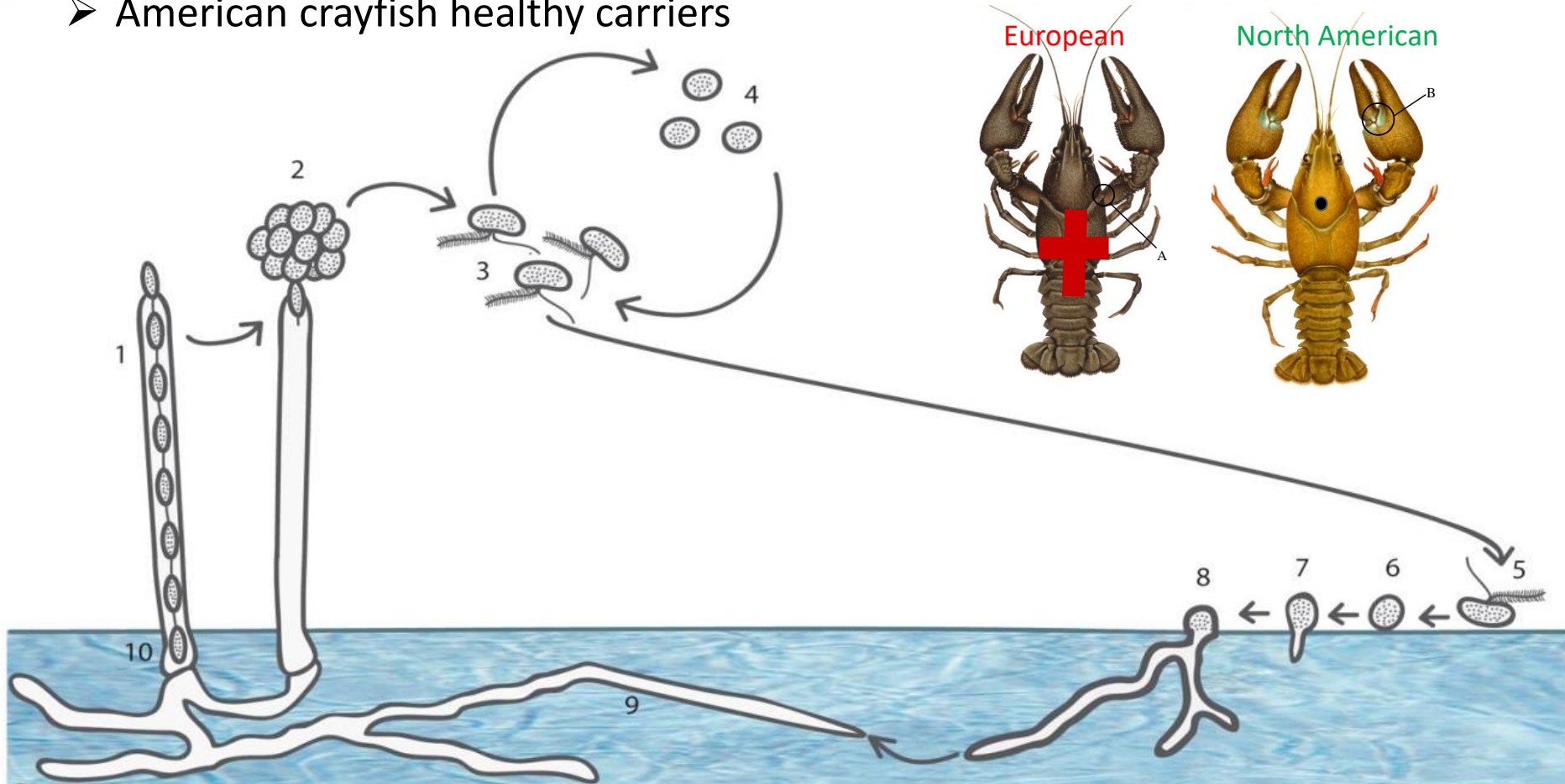
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Crayfish plague – *Aphanomyces astaci*

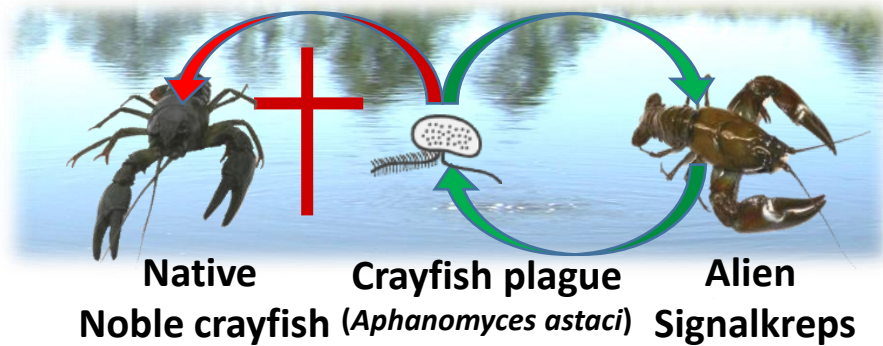
(Oomycetes » Saprolegniales » Leptolegniaceae)

- OIE listed, lethal to European crayfish
- **Norway: List 3 (national) disease**
- 100 worlds worst invasive species
- American crayfish healthy carriers



qPCR crayfish plague

From diagnostic method to eDNA monitoring tool



2012: OIE method



2009: qPCR diagnostic assay tissue samples



2011: detect in water Laboratory conditions



2012: detect in water Aquarium conditions



2014: detect in water Nature – large systems



2015: Validation during an outbreak



2016: Implementation in National surveillance



Strand et al. 2019

Strand et al. 2014

Strand et al. 2012

Strand et al. 2011

Vrålstad et al. 2009

Detection & quantification of host-pathogen eDNA in water

50 YEARS WITH IMPACT

Journal of Applied Ecology

British Ecological Society

Journal of Applied Ecology 2014, 51, 544–553

doi: 10.1111/1365-2664.12218

Detection of crayfish plague spores in large freshwater systems

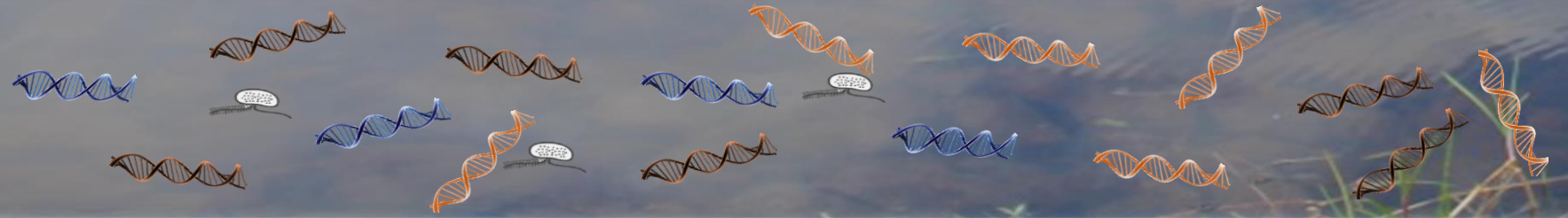
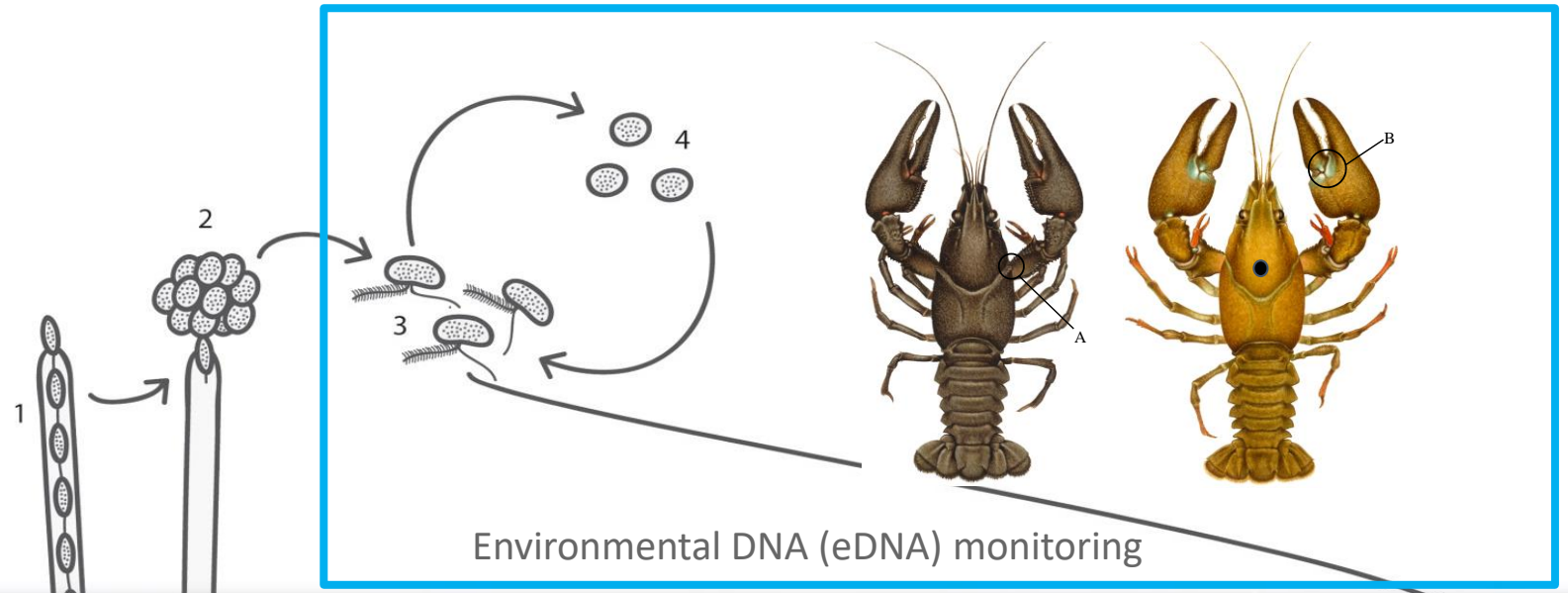
David A. Strand^{1,2}, Japo Jussila³, Stein I. Johnsen⁴, Satu Viljamaa-Dirks⁵, Lennart Edsman⁶, Jannicke Wiik-Nielsen¹, Hildegunn Viljugrein¹, Frederik Engdahl⁶ and Trude Vrålstad^{1,2*}

PLOS ONE

RESEARCH ARTICLE

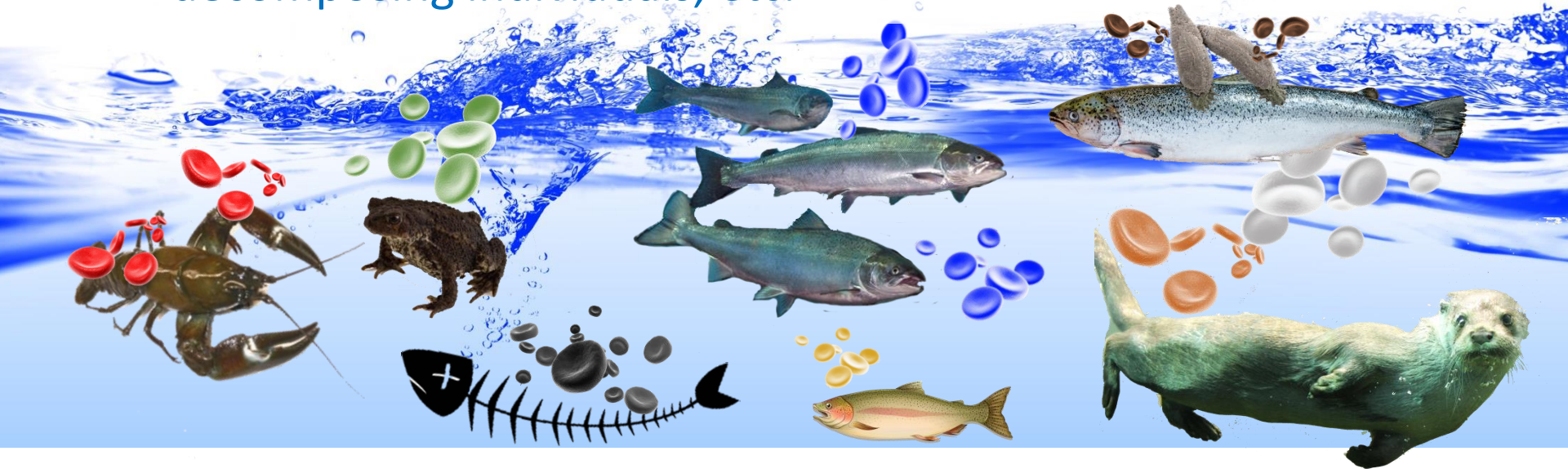
Monitoring of noble, signal and narrow-clawed crayfish using environmental DNA from freshwater samples

Sune Agersnap^{1*}, William Brenner Larsen^{1*}, Steen Wilhelm Knudsen¹, David Strand², Philip Francis Thomsen³, Martin Hesselsoe⁴, Peter Bondgaard Mortensen⁵, Trude Vrålstad², Peter Rask Møller¹



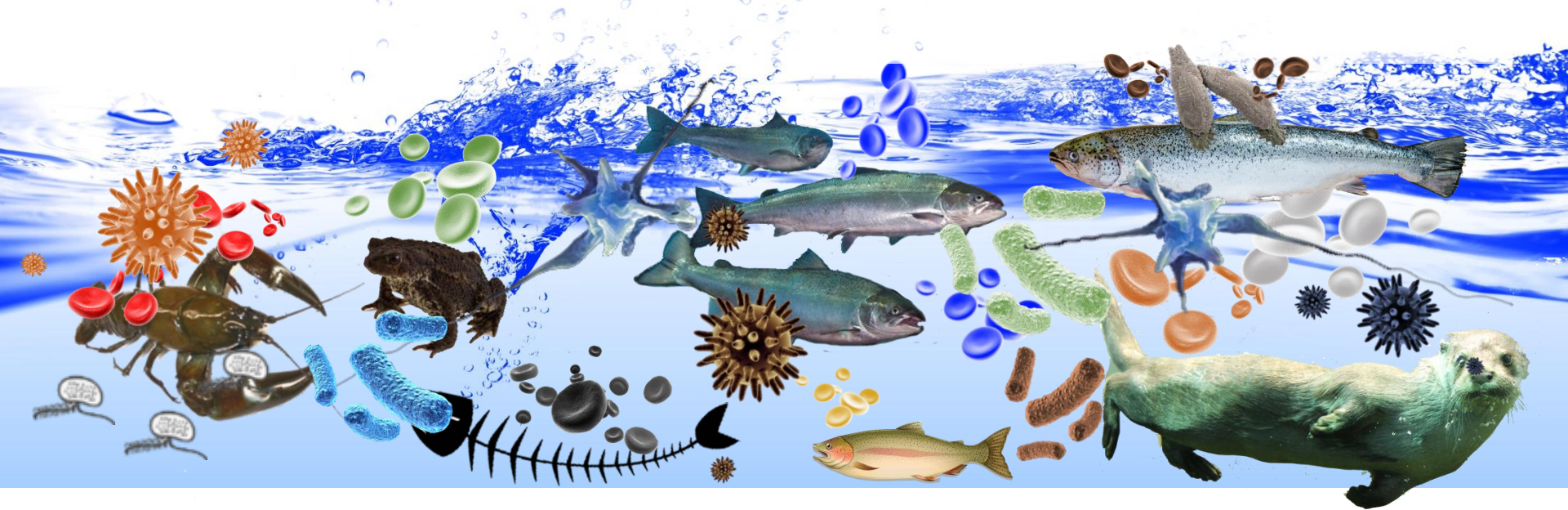
eDNA sources

- Aquatic animals shed cells with DNA to the environment through feces, skin, urine, sperm, eggs, mucus, blood, decomposing individuals, etc.



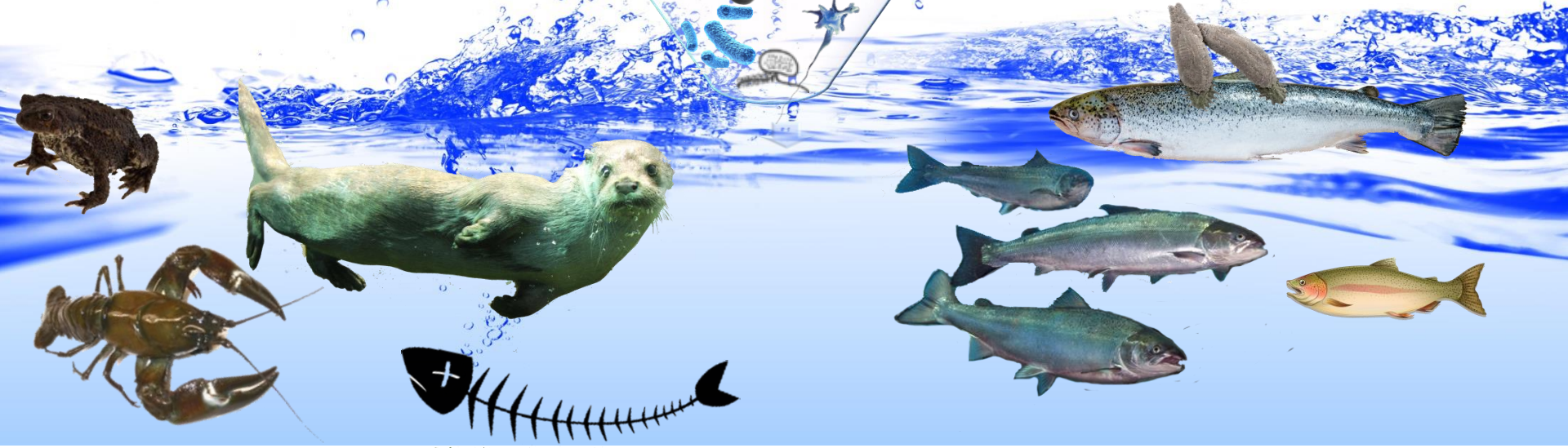
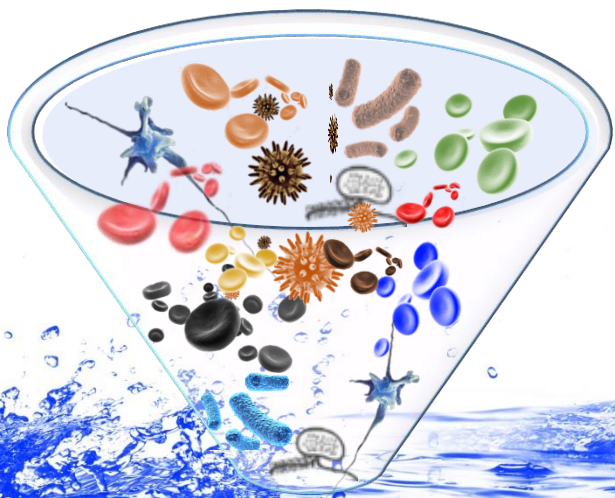
eDNA sources

- Microorganisms including pathogens lives in the environment, or are emitted from infected organisms



Concentration of cell soup using filtration

Without stressing, harming or killing

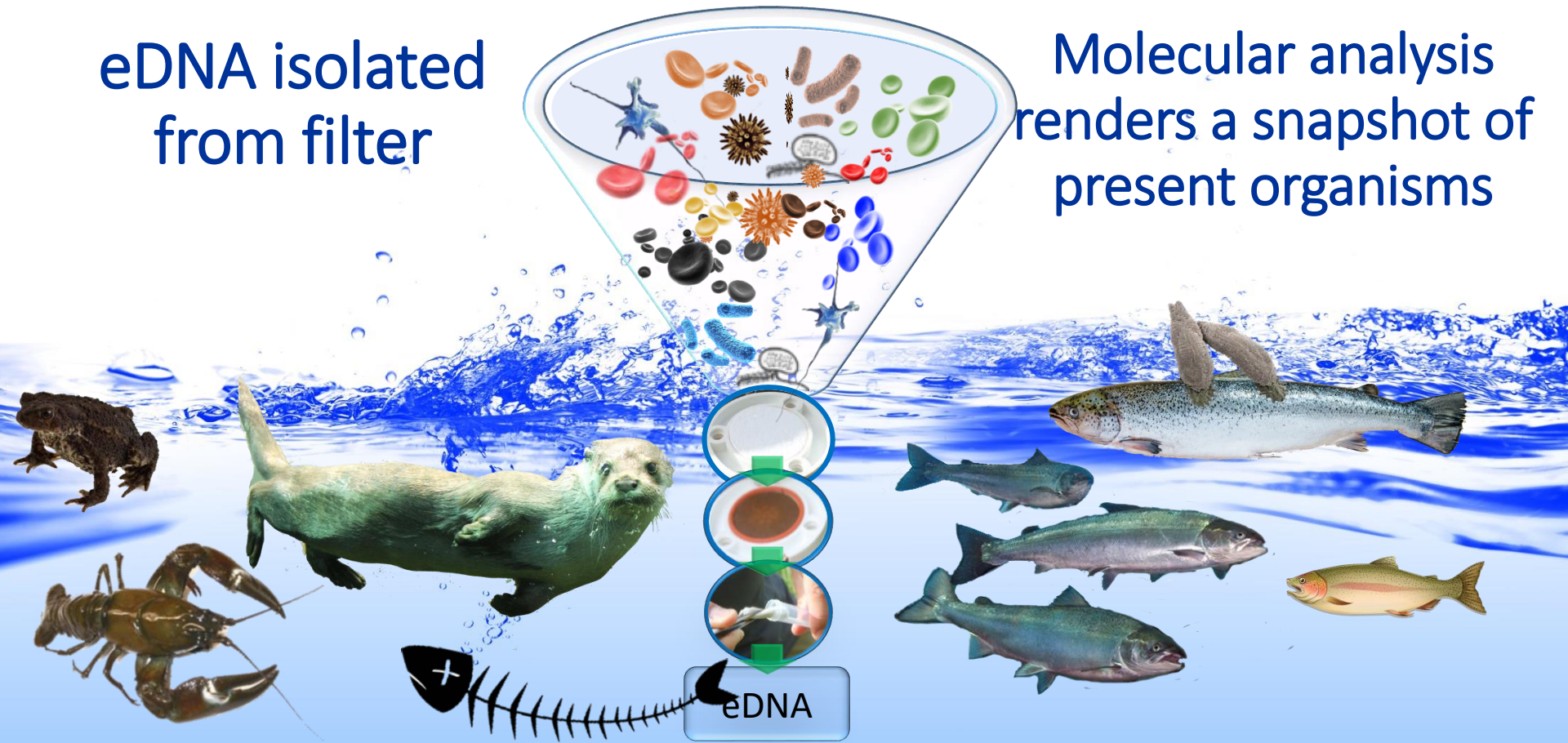


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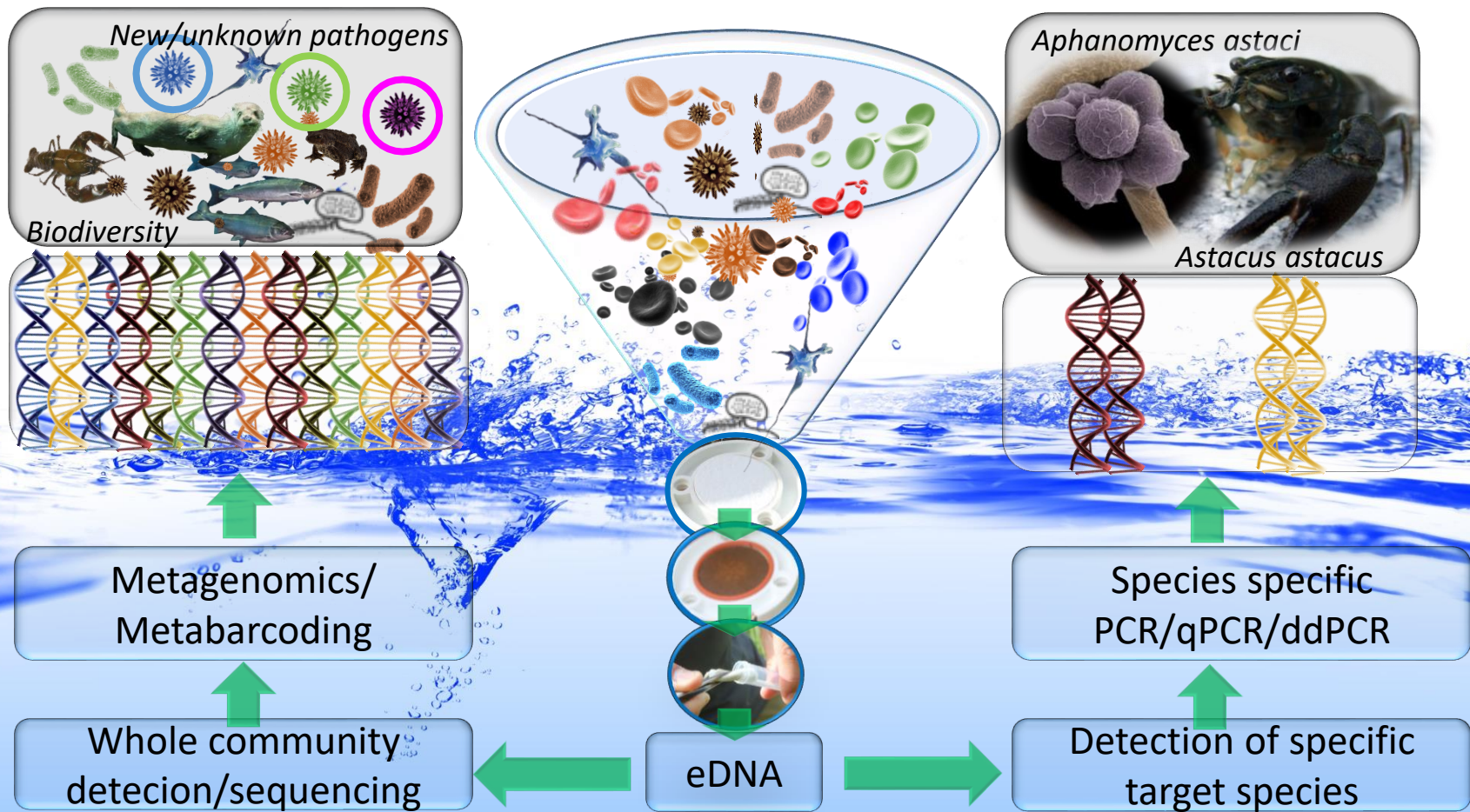
eDNA isolated
from filter

Molecular analysis
renders a snapshot of
present organisms



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eDNA sampling protocols



Collecting vessel with lead & tube



Pump, power & filtration system



Battery driven easy portable water sampler (all in one box system, expensive)



Canister measure filtered water volume (~5 L)



Disposable filter cup



Drill-powered pump (cheap)

Alternative

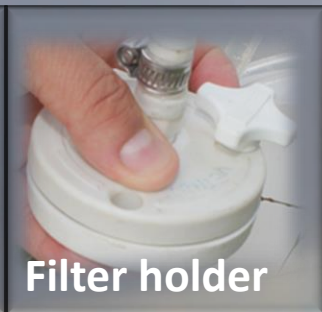
Alternative

Alternative

eDNA sampling protocols and downstream analyses



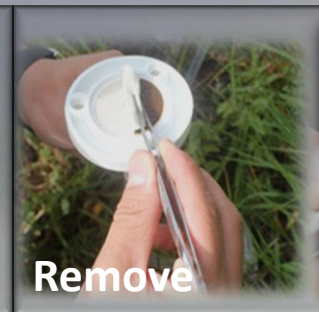
Clean filter



Filter holder



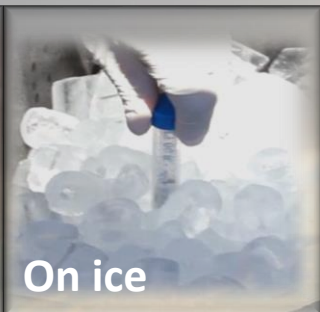
5 L later



Remove

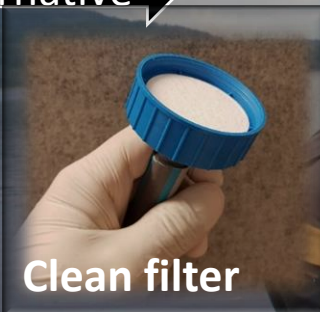


Store



On ice

Alternative



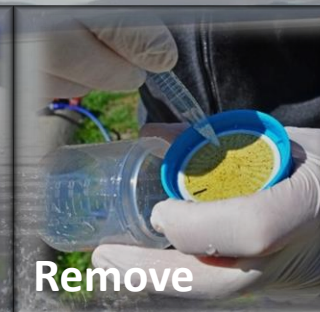
Clean filter



Add cup holder



5 L later




Remove




Store in silica

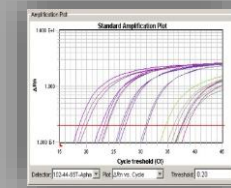
Large volume DNA extraction




qPCR



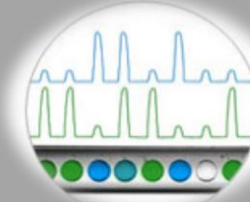
Relative quantity



ddPCR



Absolute quantity



eDNA monitoring of *A. astaci* and freshwater crayfish in a real-life situation



- Ørje locks closed since 2005
 - Stop the spread of crayfish plague and signal crayfish
- Signal crayfish moved illegally upstream Ørje locks
 - Discovered during noble crayfish surveillance in 2014
 - **The onset of a crayfish plague outbreak unavoidable**







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RESEARCH ARTICLE

Journal of Applied Ecology 

Monitoring a Norwegian freshwater crayfish tragedy: eDNA snapshots of invasion, infection and extinction

David A. Strand^{1,2}  | Stein Ivar Johnsen³ | Johannes C. Rusch^{1,4}  | Sune Agersnap^{5,6}  | William Brenner Larsen⁵ | Steen Wilhelm Knudsen⁵  | Peter Rask Møller⁵  | Trude Vrålstad¹ 

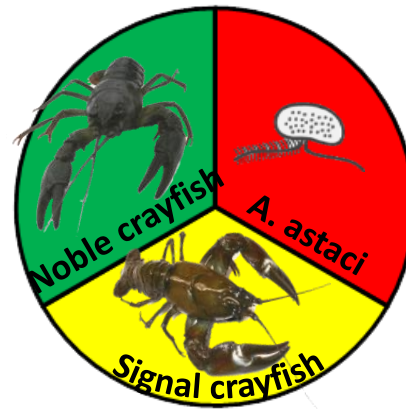


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Comparing methods during an outbreak

- Compare conventional surveillance with eDNA monitoring
- National surveillance noble crayfish = trapping
- National surveillance crayfish plague = cage trials



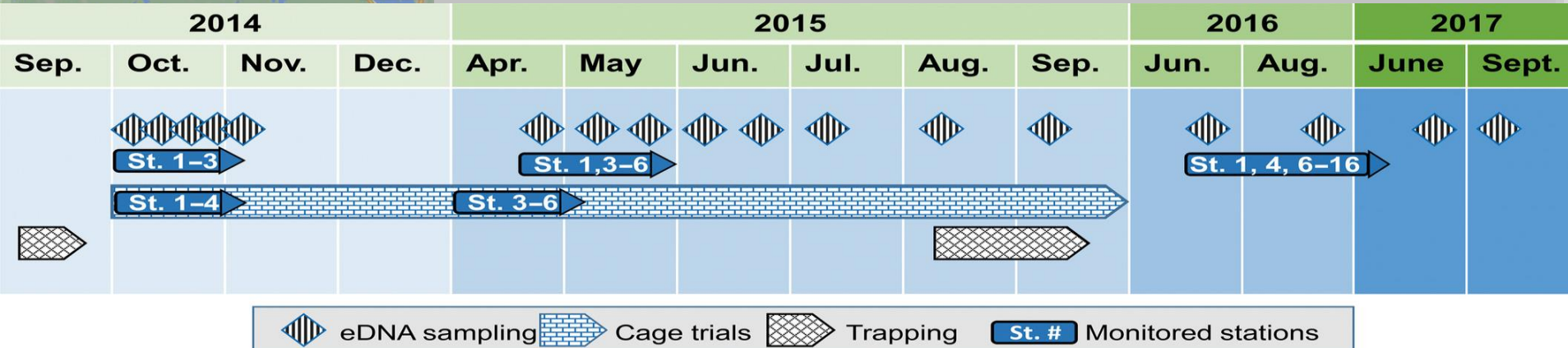
eDNA monitoring

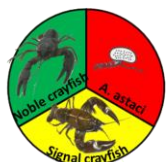


Cage trials



Trapping (CPUE)





eDNA monitoring



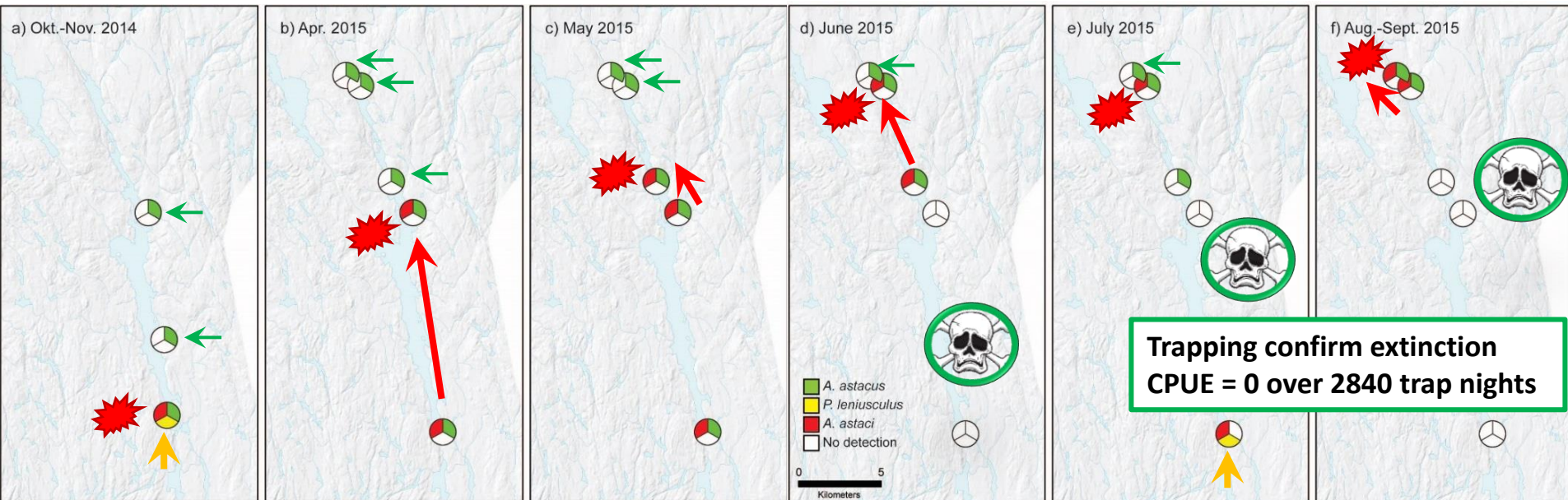
Cage trials



Trapping (CPUE)

Comparing methods during an outbreak

- The crayfish plague outbreak started late in 2014.
- *A. astaci* eDNA detected 3 weeks before mortality of crayfish observed in the cages
- Upstream spread also during winter
- After one year, 23 km spread
- Noble crayfish died in the cages and lake – and eDNA of both targets vanished
- A huge trapping effort with 0 noble crayfish confirm local extinction
- Detect signal crayfish at low densities (~ 0.12 CPUE) – but not every time



Oktober

April

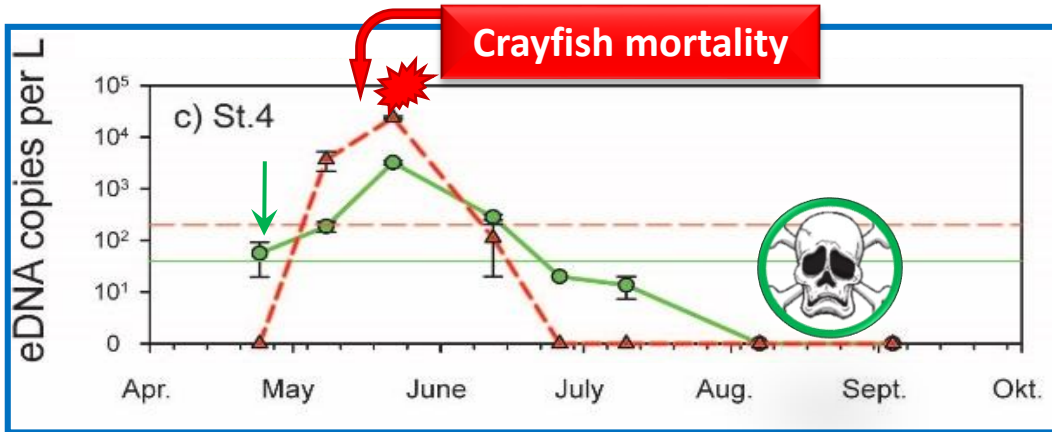
Mai

Juni

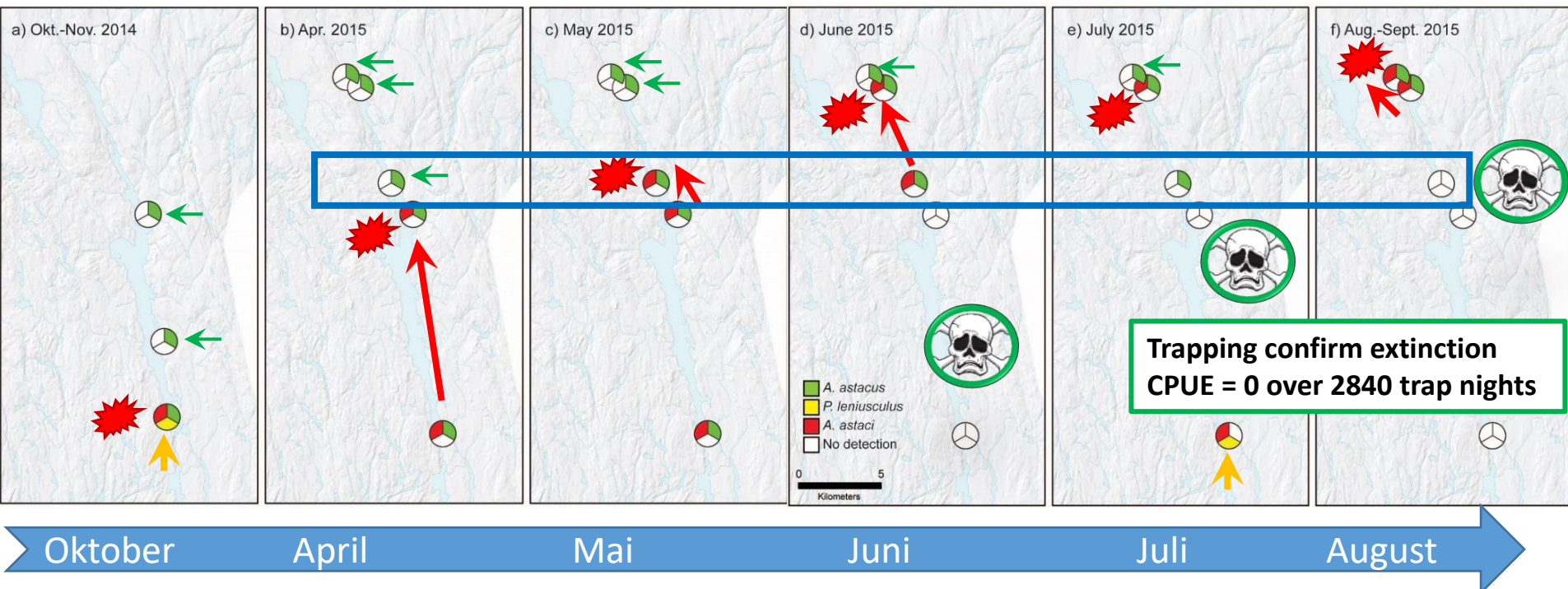
Juli

August

eDNA fluctuations - from infection to extinction



- First only eDNA of noble crayfish.
- Then *A. astaci* detectable at low levels – then increased
- Parallel trend for noble crayfish eDNA
- Then eDNA of both targets disappeared, reflecting the local extinction



eDNA snapshots of a crayfish plague outbreak

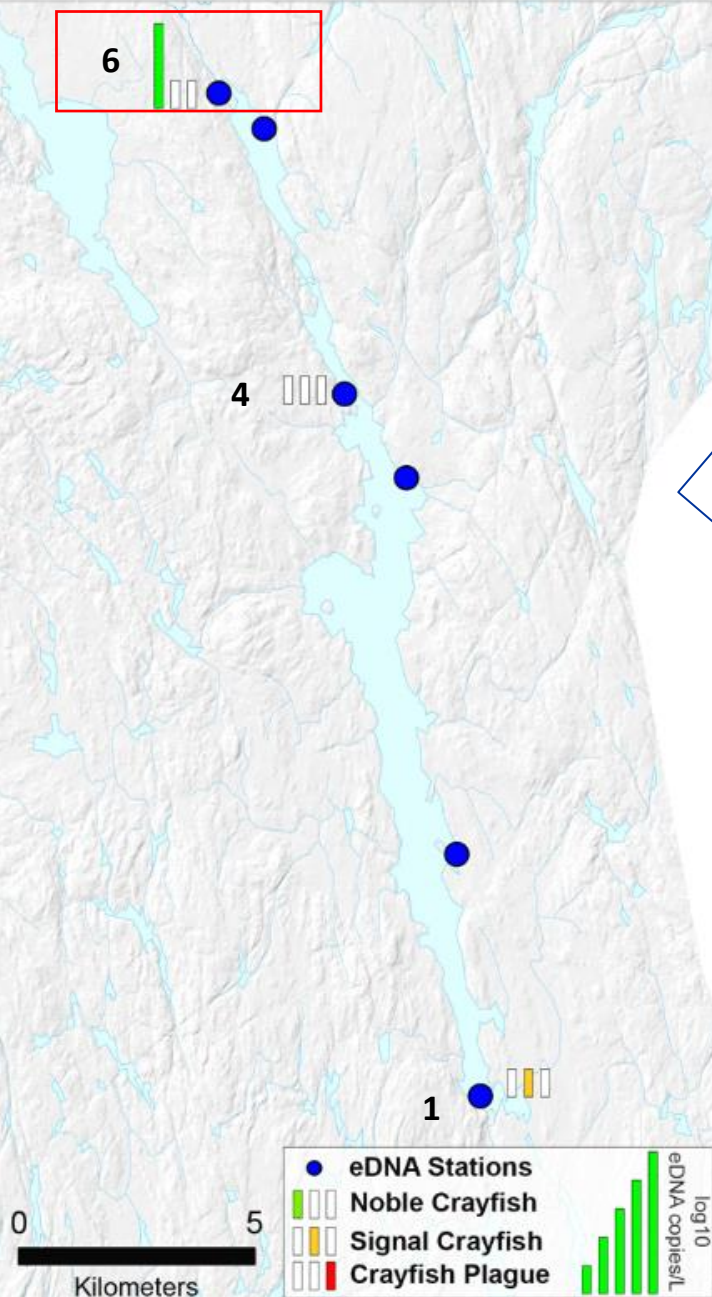
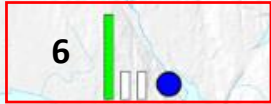


- 2014
 - eDNA of all targets at the water locks
 - Upstream spread under the ice
- 2015
 - Outbreak spreads steadily
 - eDNA increase during outbreak – then disappear
- 2016
 - The outbreak front stagnated and burnt out
- 2017
 - *A. astaci* only seen in the signal crayfish location
 - Noble crayfish eDNA remain where the outbreak burnt out
- See the time-lapse here:
<https://appliedecologistsblog.com/2019/06/07/edna-snapshots-of-invasion/>

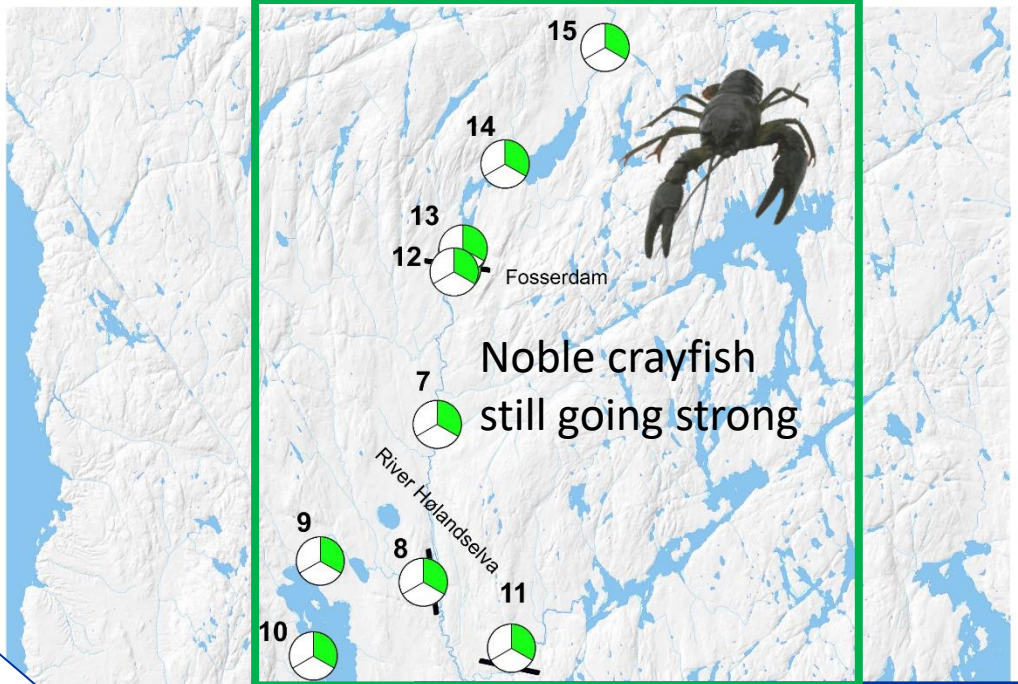


eDNA snapshots of a crayfish plague outbreak

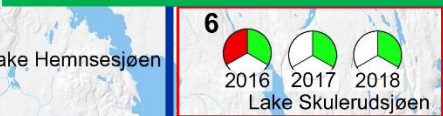
Sept. 2017



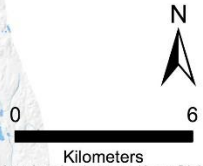
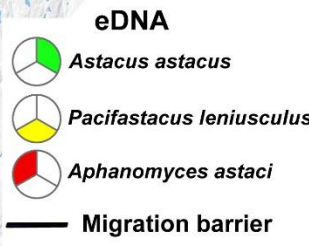
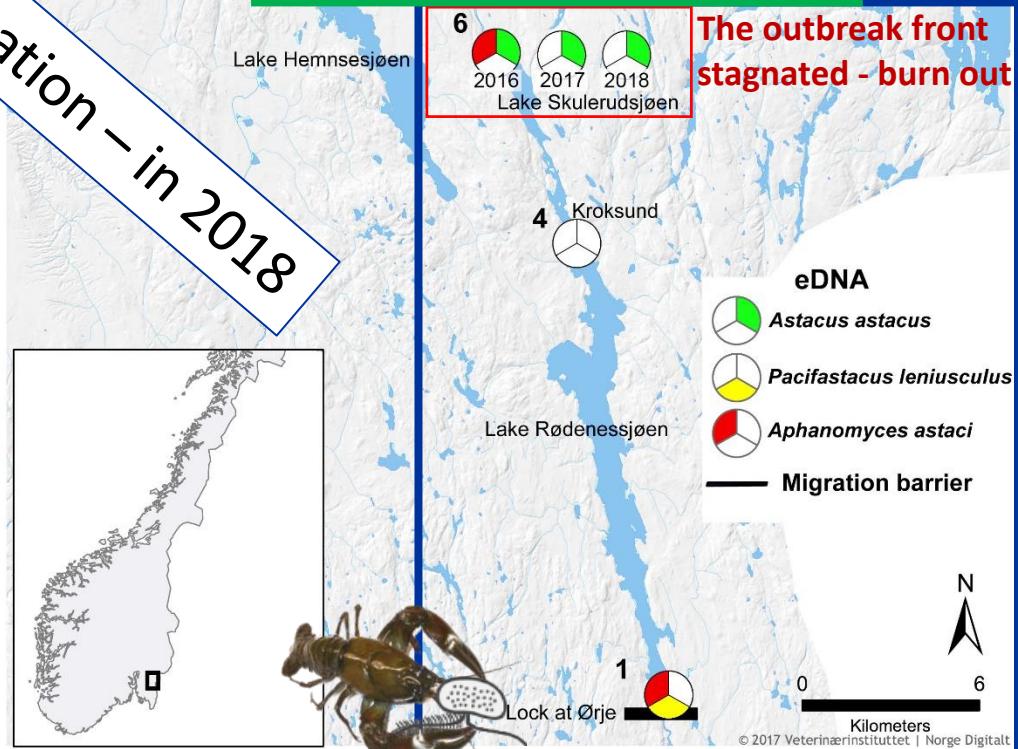
Continuation – in 2018



Noble crayfish still going strong



The outbreak front stagnated - burn out



The surveillance programme for *Aphanomyces astaci* in Norway 2016



The surveillance programme for *Aphanomyces astaci* in Norway 2017



eDNA implemented in Norwegian surveillance

- 2016: eDNA supplement cage experiments in *A. astaci* surveillance
- 2017: eDNA replace cage experiments in *A. astaci* surveillance
- 2018: Joint use of eDNA results *A. astaci* and crayfish surveillance programs
- Continues in 2019...

The surveillance programme for *Aphanomyces astaci* in Norway 2018



Coordination work and joint results



1590

National surveillance of noble crayfish, and spread of signal crayfish

Stein I. Johnsen, David A. Strand, Johannes Rusch & Trude Vrålstad

NINA Rapport



Samarbeid og kunnskap for framtidens miljøløsninger



eDNA study in Czech Republic (2017-2019)



Rusch et al. *in prep*

Evaluate the presence of *A. astaci* and crayfish in waterbodies within Czech Republic
Collaboration with Prof. Adam Petrusek, Charles University Prague



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Mobile technology and on-site detection of crayfish plague



https://www.youtube.com/watch?v=flvIH_Zj9aM&feature=youtu.be



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Summary - eDNA monitoring



- An efficient and rapid monitoring tool – but still only a snapshot
- Fluctuations of eDNA amount can reflect stages of infection – outbreak - extinction
- Detect the infectious spores in the water earlier than caged crayfish
- Simultaneous detection of crayfish eDNA – extra confirmation for little extra cost
- Well suited for
 - Early warning of invasion or infection
 - Risk assessments / Habitat evaluation / Surveillance of pathogen/host
- Non-invasive and animal welfare friendly
- Has replaced the cage trials with live crayfish (“canaries in a coalmine”)
- What is next?



Automated continuous eDNA monitoring?



Acknowledgements

- TARGET (NFR-293407)



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Norwegian Veterinary Institute

- National surveillance *Aphanomyces astaci*



- National surveillance program Noble crayfish



- Co-authors and collaborators

- David Strand, Johannes Rusch, Elin Rolén, Hildegunn Viljugrein, NVI



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- Tom Andersen, UiO



UiO • University of Oslo

- Stein Ivar Johnsen, NINA



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- Peter Rask Møller, Sune Agersnap, Steen Knudsen, UoC

UNIVERSITY OF
COPENHAGEN



- Adam Petrusek, CUNI



Charles University, Prague



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