

Investigating disease dynamics in shore crabs, *Carcinus maenas*, including two new Haplosporidia species.

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Introduction



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Common shore crab
Carcinus maenas



Introduction



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Parasitic dinoflagellate *Hematodinium* sp.



Brown crab (*Cancer pagurus*)



Velvet swimming crab (*Necora puber*)



Tanner crab (*Chionoecetes bairdi*) and snow crab (*C. opilio*)



Norwegian lobster (*Nephrops norvegicus*)



Blue crab (*Callinectes sapidus*)



the sand crab *Portunus pelagicus*





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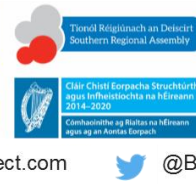
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Questions:

- Where are the reservoirs of shellfish diseases in the aquatic environment?
- Do common shore crabs, *Carcinus maenas*, harbour pathogens potentially dangerous to crustaceans of commercial interest?
- Can we 'track' pathogens, such as *Hematodinium*, in the water column and into the host using eDNA approaches?

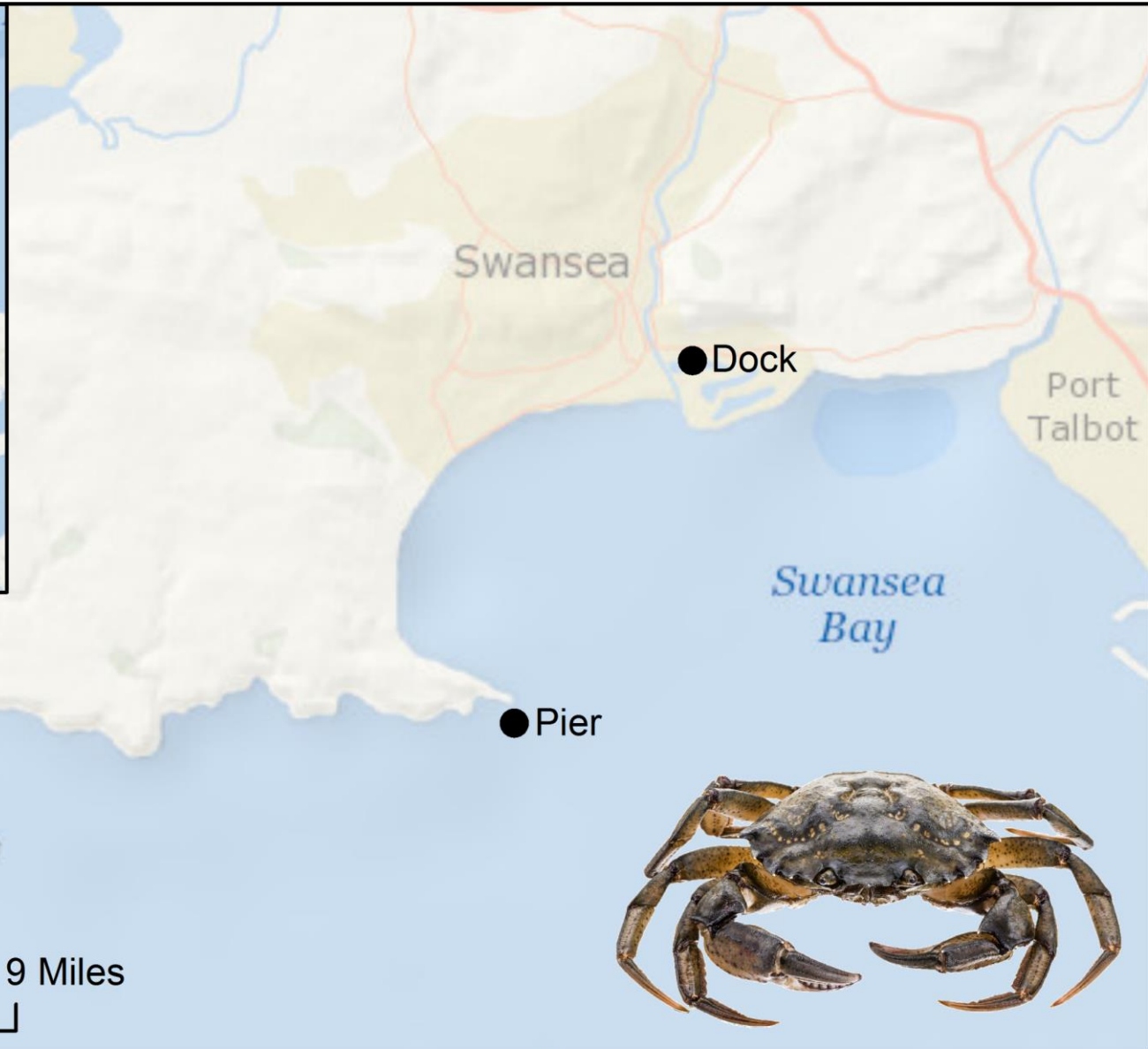
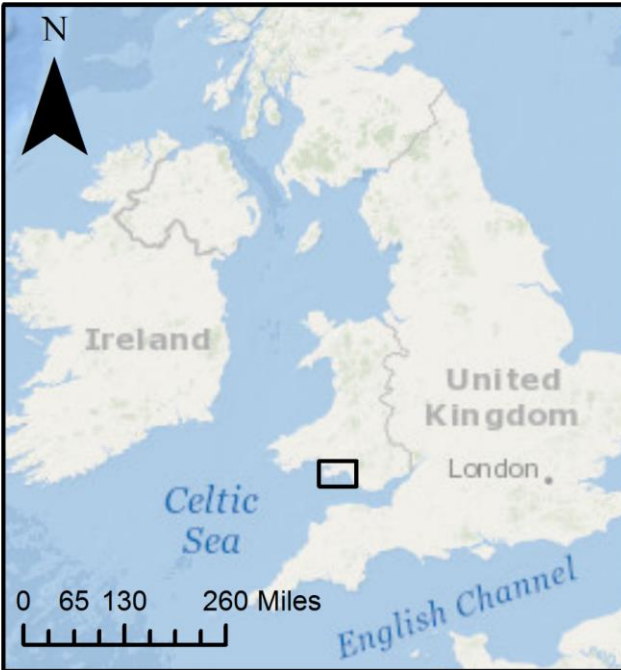
Location



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Methods



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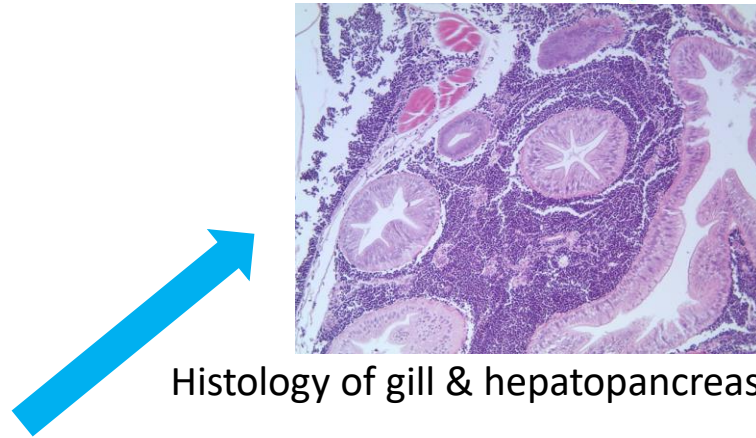
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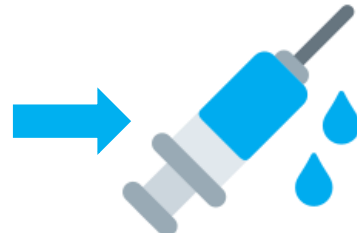
Crabs



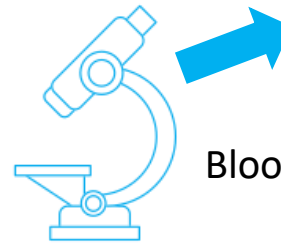
50/ month / location



Histology of gill & hepatopancreas



Haemolymph



Blood smears



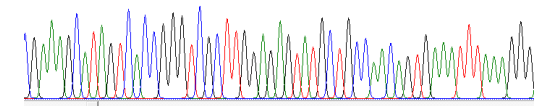
Seawater



Filtration

DNA extraction and PCR with *Hematodinium* specific primers

120 160 170 180 190 200
G A A A G C A T A G T C A C C G G G T C G C T T T G G A G A G T A T C C C A A C A G T G A A A T T T A A G G G



Positives sequenced via Sanger and deposited in GenBank

1 L / 3 replicates / month / location

Results - PCR



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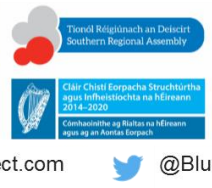
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Tested effects of the following on *Hematodinium* presence:

- Season (winter [Dec, Jan, Feb], spring [Mar, Apr, May], summer [Jun, Jul, Aug], autumn [Sept, Oct, Nov])
- Size (Carapace width, CW [continuous number])
- Sex (male or female)
- Colour (green, yellow or orange)
- Pigment loss (0 or 1)
- Haemolymph opacity (milky or clear)
- Fouling (presence of epibionts, 0 or 1)
- Limb loss (0 or 1)
- Location (Pier or Dock)

Davies et al. (2019) *Parasites & Vectors*
doi: [0.1186/s13071-019-3727-x](https://doi.org/10.1186/s13071-019-3727-x)

Results - PCR



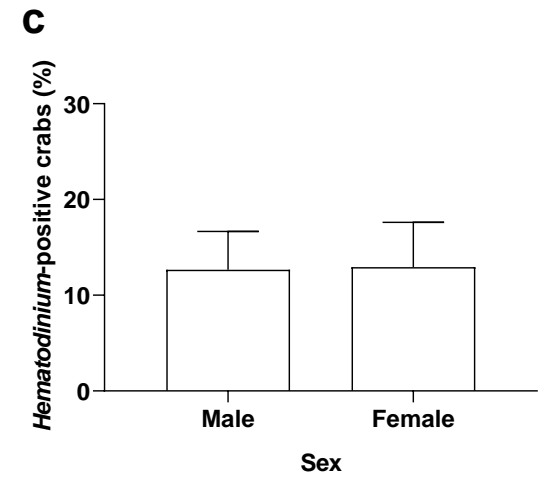
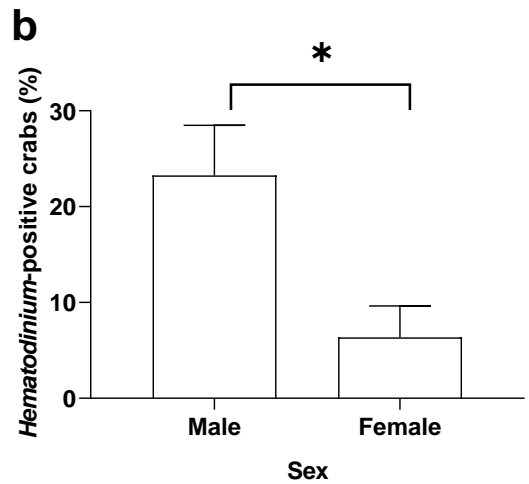
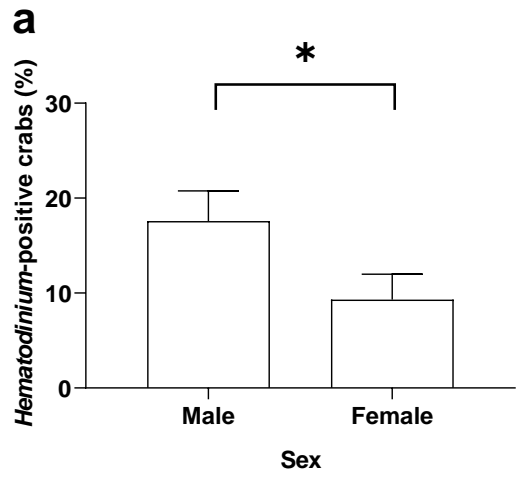
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Total
13.6% infected

Dock
14.4% infected

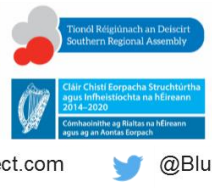
Pier
12.8% infected



Sex

Males nearly twice as likely to become infected overall.

Results - PCR



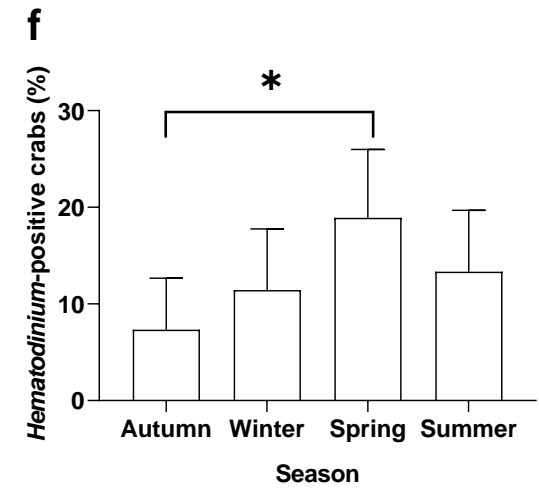
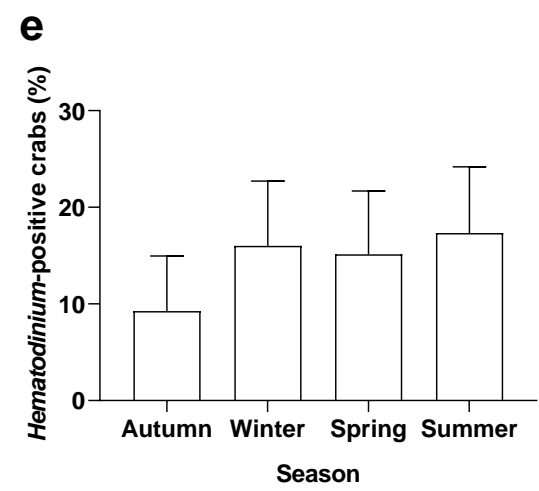
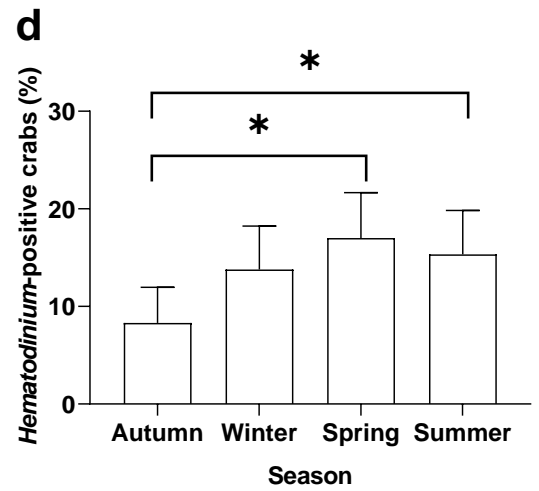
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Total

Dock

Pier

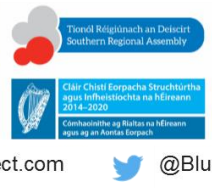


Season

Crabs in spring and summer more likely to be infected.

Davies et al. (2019) *Parasites & Vectors*
doi: [0.1186/s13071-019-3727-x](https://doi.org/10.1186/s13071-019-3727-x)

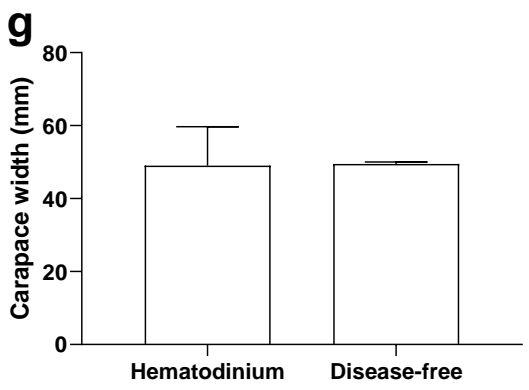
Results - PCR



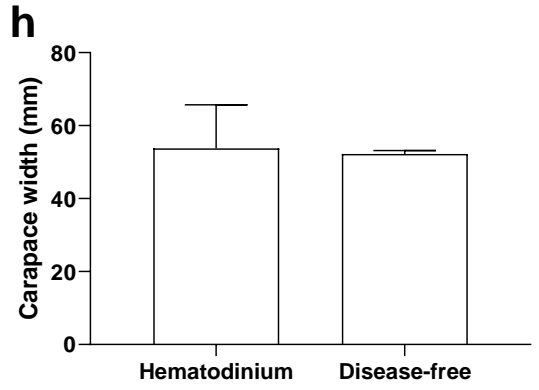
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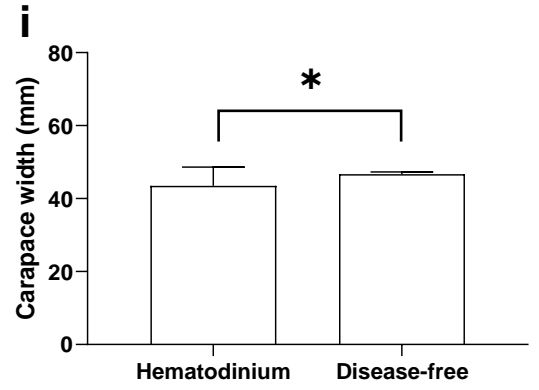
Total



Dock



Pier



Size

Smaller crabs more likely to be infected, in **Pier** location only.

Results



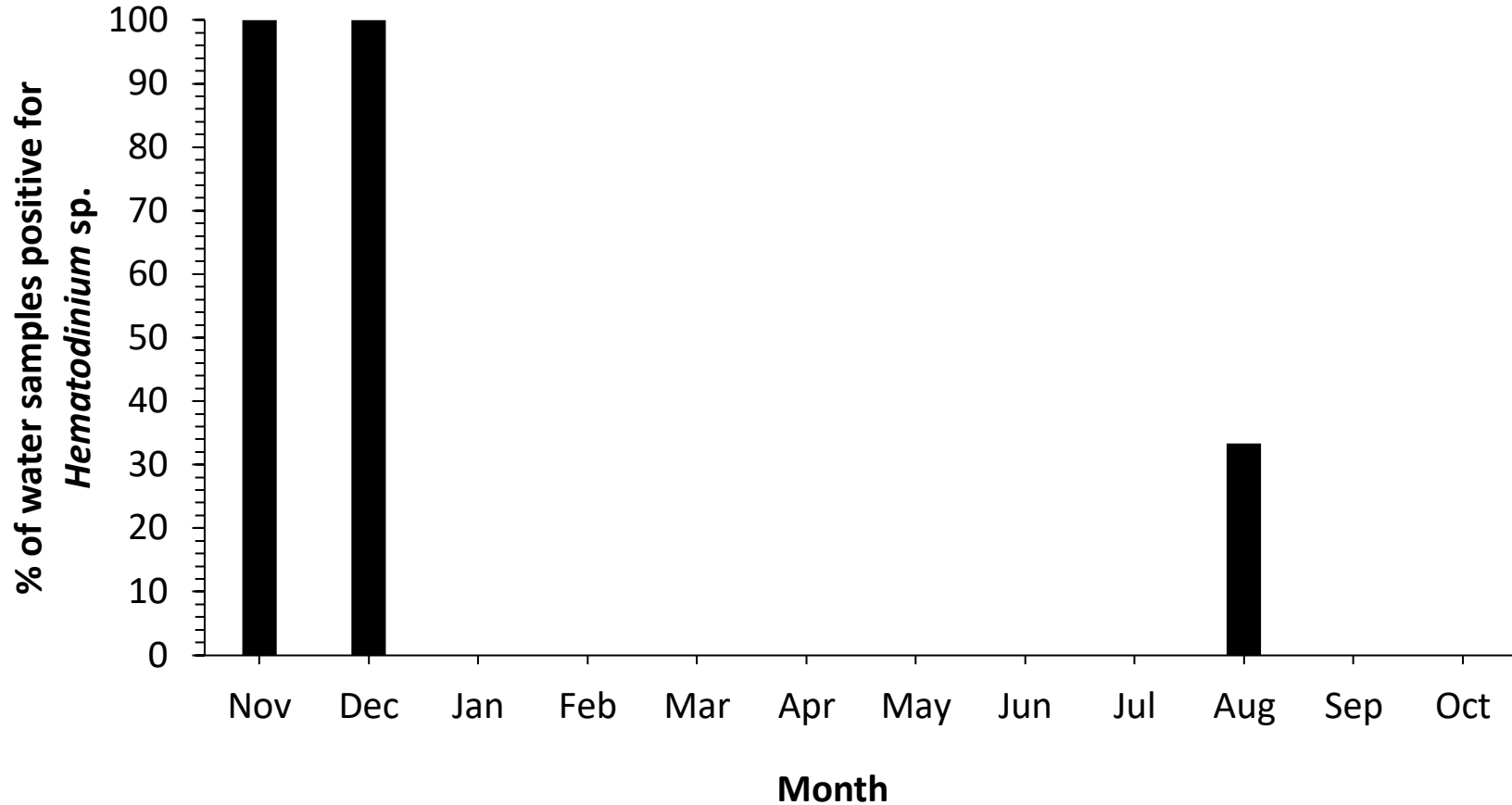
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Water eDNA

Hematodinium sp. ONLY found in the **Pier**, not the Docks.

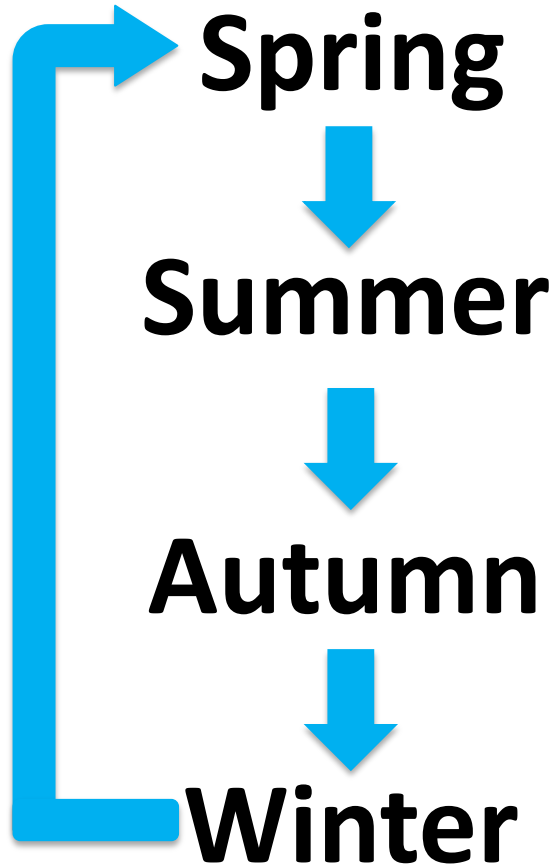


Davies et al. (2019) Parasites & Vectors
doi: [0.1186/s13071-019-3727-x](https://doi.org/10.1186/s13071-019-3727-x)

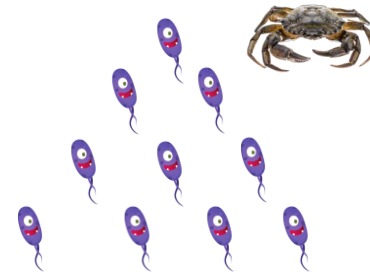



Life Cycle


Possible *Hematodinium* sp. life cycle?



Peak **prevalence** of *Hematodinium* sp. in crabs
- Multiplication of *Hematodinium* sp. in crab tissues



Highest **severity** of *Hematodinium* sp. infection in gills, hepatopancreas and haemolymph of crabs 

Severely infected, moribund crabs release infective stage dinospores into water
Infection of new crabs
Hematodinium sp. found in **eDNA** from water column 

Co-infections



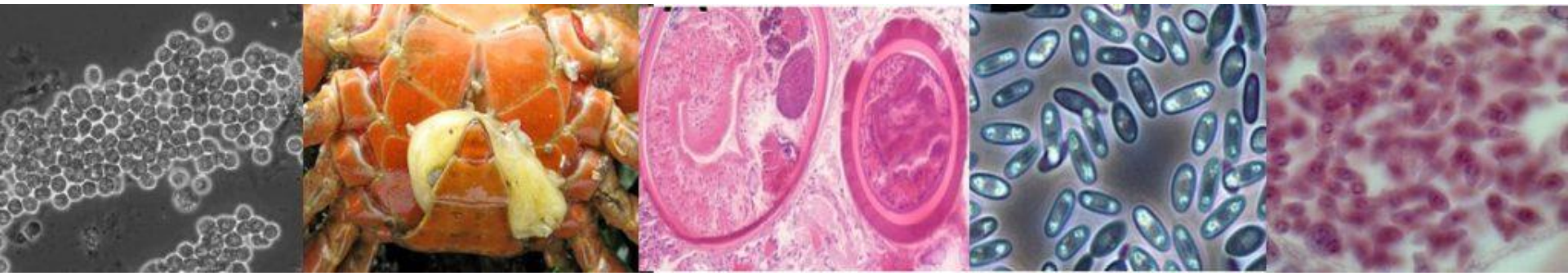
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Pathogen	<i>C. maenas</i>		Seawater eDNA	
	Dock	Pier	Dock	Pier
<i>Haplosporidia</i>	X	✓	✓	✓
<i>Microsporidia</i>	✓	X	✓	X
Mikrocytids	X	X	✓	✓
Paramyxids	X	X	X	✓
<i>Vibrio</i> spp.	✓	✓	✓	✓
Fungal species	X	✓	✓	✓
Trematode parasites	X	✓	NA	NA
<i>Sacculina</i> sp.	✓	X	NA	NA

Davies et al. (2021) Preprint. BioRxiv DOI: [10.1101/2021.05.26.445743](https://doi.org/10.1101/2021.05.26.445743)



Co-infections

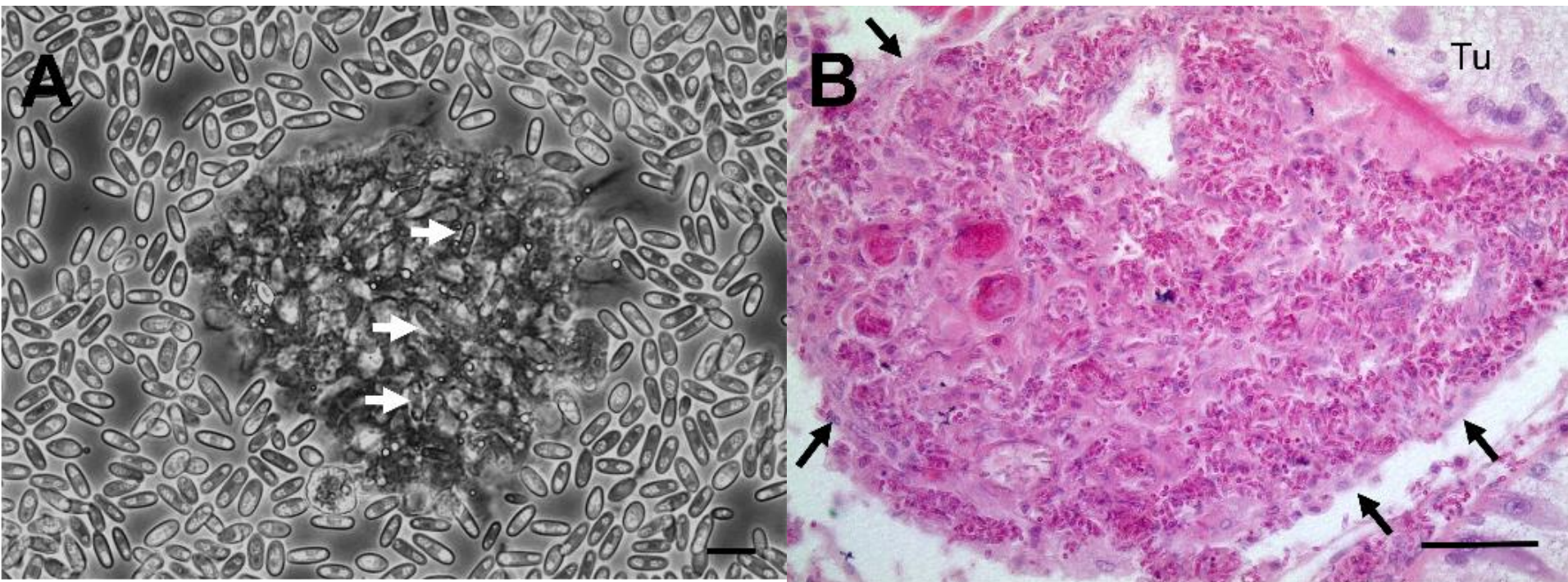


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Fungi, or a novel mycosis?



Fungi

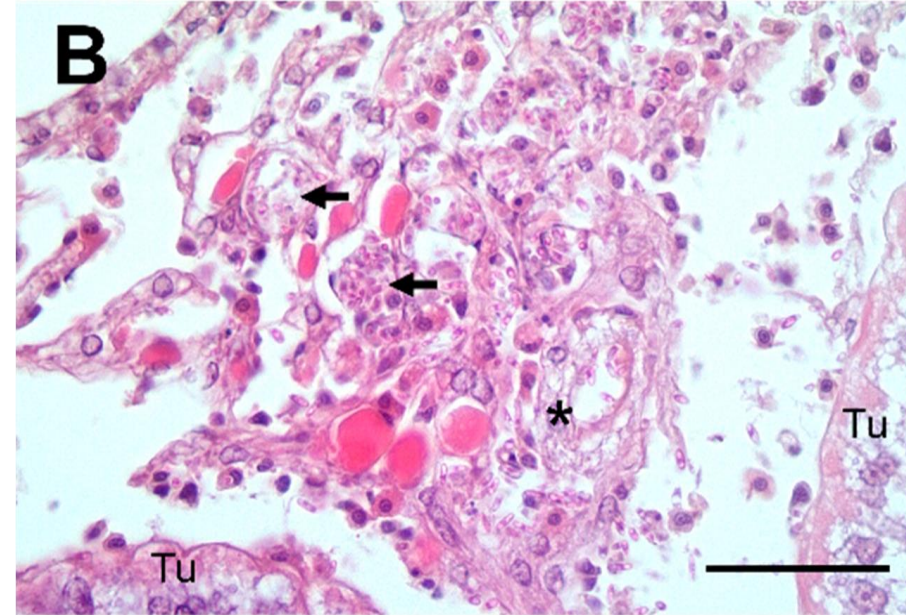


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- 87 --- Hypocrea lutea AB027338
- Verticillium incurvum AF339600
- Cordyceps irangiensis DQ522556
- marine fungal sp. FCAS20 GQ120173
- Fusarium solani AB473810
- Simplicillium lamellicola AF339601
- 73 --- marine fungal sp. FCAS129 GQ120159
- Simplicillium lanosoniveum HQ232185
- 87 --- Hirsutella sp. EF469125
- Ophiocordyceps stylophora DQ522552
- **Hypocreales sp. crab fungus JX480545**
- Ophiocordyceps entomorrhiza EF468954
- Ophiocordyceps gracilis EF468955
- Elaphocordyceps ophioglossoides JN941736
- Ascomycete sp. MV 19C marine EF638698
- Balansia henningsiana AY545723
- Claviceps purpurea AF543765
- Pochonia bulbillosa AF339591
- Paecilomyces marquandii HQ263110
- Cordyceps brongniartii JN941760
- Cordyceps pruinosa AB044629
- Cordyceps militaris AB070374
- Ascomycota sp. MV13 marine EU887738
- Cordyceps pseudomilitaris JN941748
- Cordyceps tuberculata GQ249960

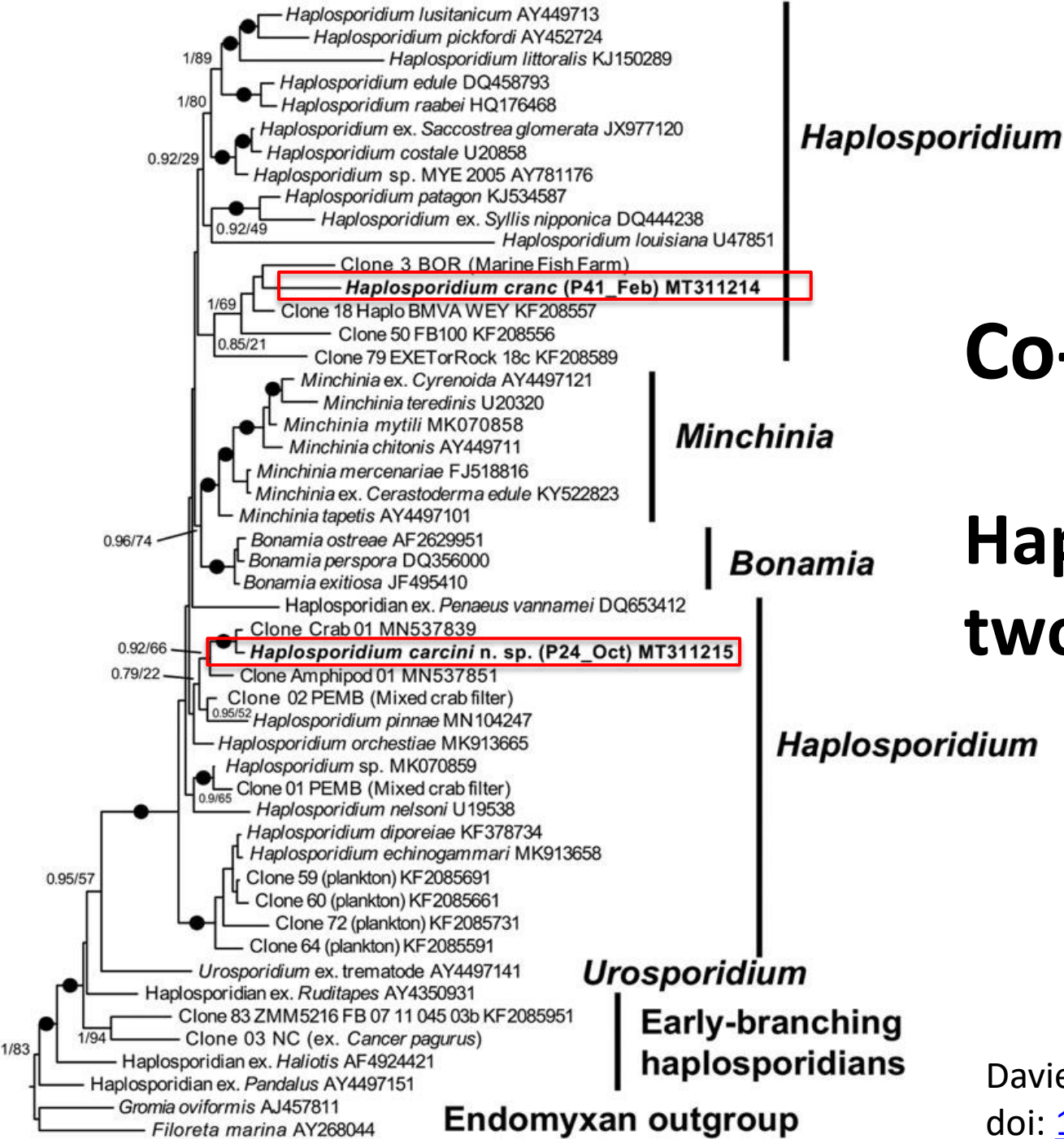


- 80 --- KY924812.1 Ophiocordyceps sp.
- KY924813.1 Ophiocordyceps sp.
- 90 --- MG245625.1 Uncultured fungus clone JPNQ46G03DDZ5U
- KY924809.1 Ophiocordyceps sp.
- 96 --- AB027374.1 Ophiocordyceps sobolifera
- Fusarium oxysporum MG727665
- 98 --- LS963216.1 Uncultured fungus
- LS958246.1 Uncultured fungus
- 99 --- KF016996.1 Aschersonia narathiwatensis strain BCC49498
- KF016997.1 Aschersonia narathiwatensis strain BCC49499
- KC713632.1 Aschersonia sp. BCC49496
- KC713631.1 Aschersonia sp. BCC49495
- 91 --- AB968397 Ophiocordyceps coenomyia
- NR 147509.1 Ophiocordyceps coenomyia NBRC 108993
- 83 --- **MT000101 P50 May**
- **MT000100 P5 Dec**
- **MT000102 P21 Aug**
- 72 --- HM119586.1 Ophiocordyceps gracilis
- HM142942.1 Ophiocordyceps gracilis
- 82 --- KY321861.1 Ophiocordyceps gracilis
- KC963033.1 Cordyceps sp. SCALT1007-002
- AJ786565.1 Cordyceps gracilis
- AJ786564.1 Cordyceps gracilis
- AJ786563.1 Cordyceps gracilis



Tree scale: 0.1

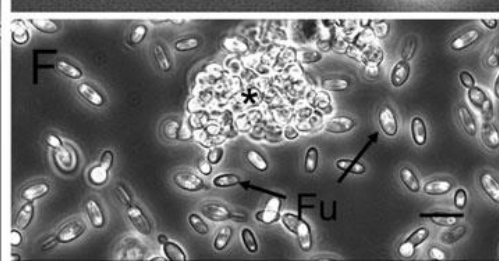
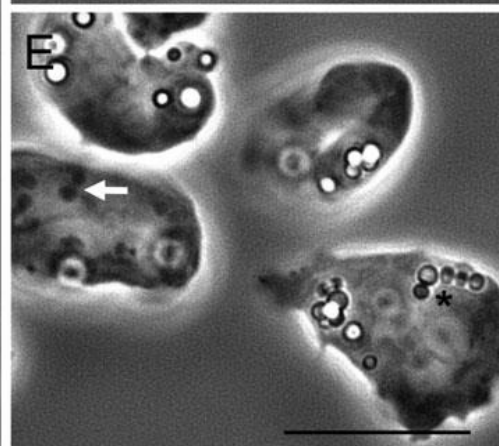
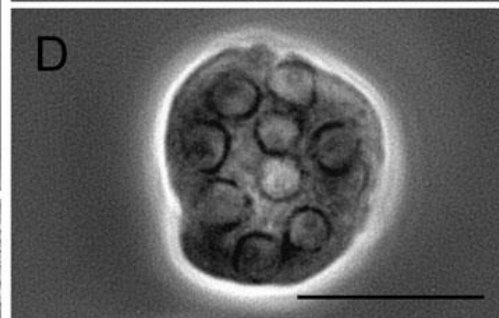
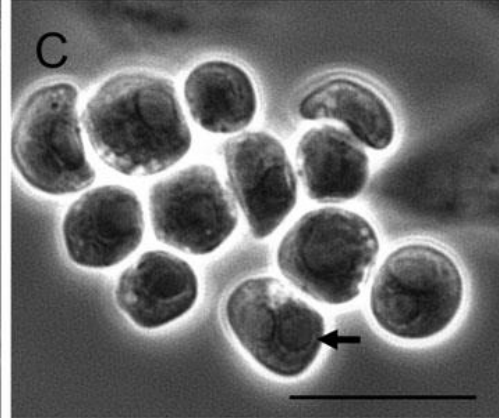
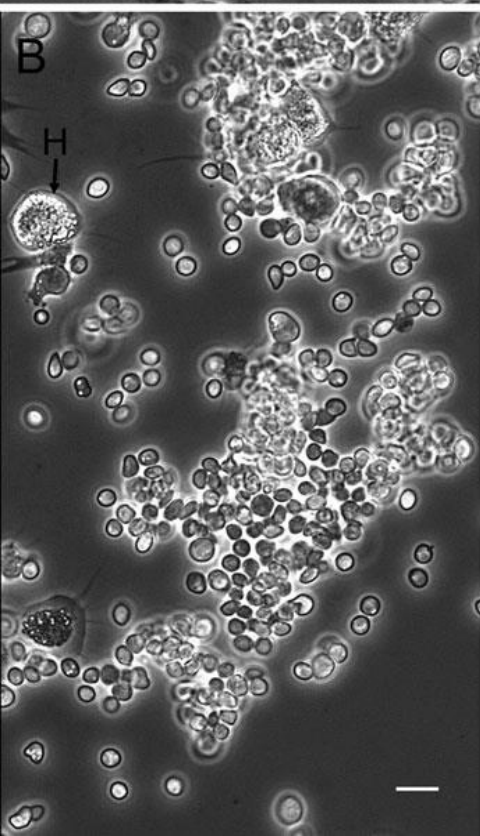
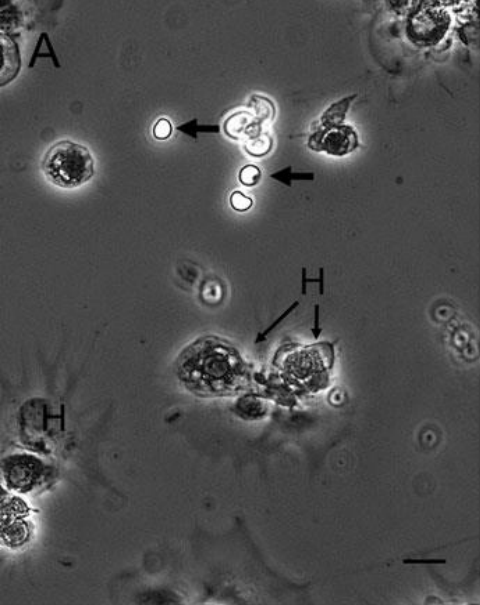
Davies et al. (2020) *Pathogens*
doi:[10.3390/pathogens9060462](https://doi.org/10.3390/pathogens9060462)



Co-infections

Haplosporidia, two new species!

Davies et al. (2020) *Parasitology*
doi: [10.1017/S0031182020000980](https://doi.org/10.1017/S0031182020000980)

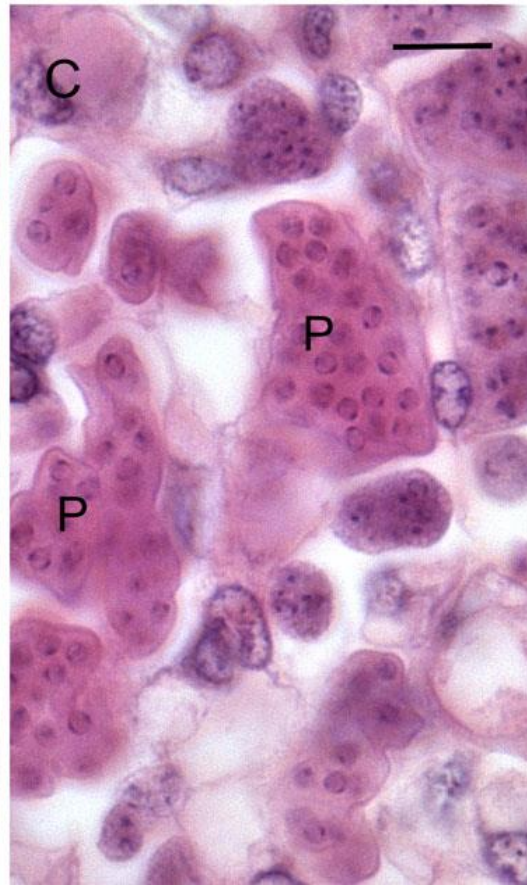
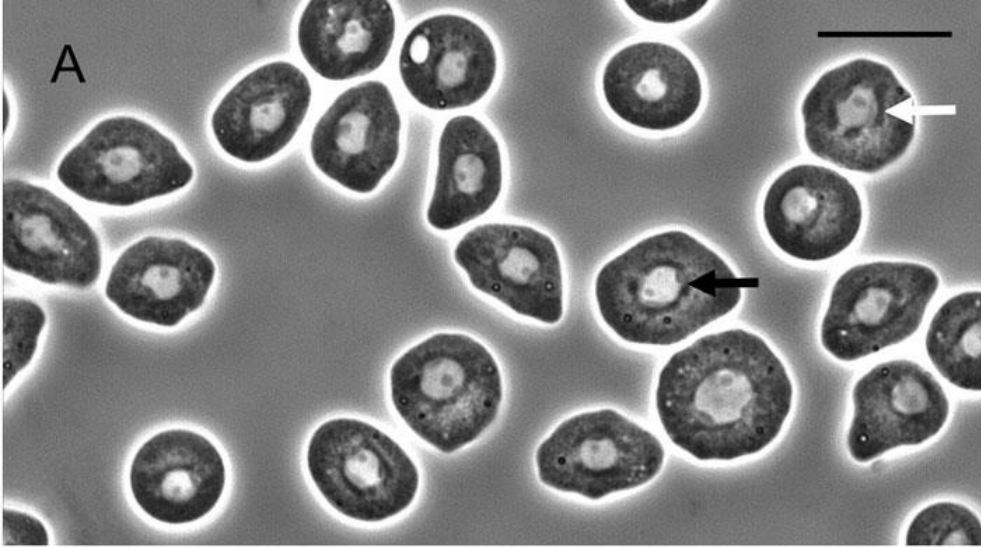


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Haplosporidium carcini

- (A). Apparent low-grade infection with adherent haemocytes (H) and non-adherent, refractile haplosporidia (unlabelled arrows).
- (B). Higher severity infection with numerous uninucleate parasites and smaller number of haemocytes (H).
- (C). Uni- and bi-nucleate (arrow) forms of parasite.
- (D). Multinucleated plasmodium.
- (E). Spread cells of unknown origin with cytoplasmic refractile bodies (*) and bacteria-like structures (arrow).
- (F). Coinfection of *H. carcini* (*) and an unidentified yeast-like fungus (Fu). Scale bars = 10 μ M.

Davies et al. (2020) *Parasitology*
doi: [10.1017/S0031182020000980](https://doi.org/10.1017/S0031182020000980)



Haplosporidium cranc

(A). Mononucleated haplosporidians in the haemolymph of an infected crab. Note unusual chromatin arrangement (unlabelled arrows).

(B). Uninucleate haplosporidians in the terminal vessel of a gill lamella. Note prominent chromatin blocks in these cells (unlabelled arrow).

(C). Multinucleated plasmodial forms (P) of this parasite in the interstitial space of the hepatopancreas. Scale bars = 10 μ M.

Co-infections



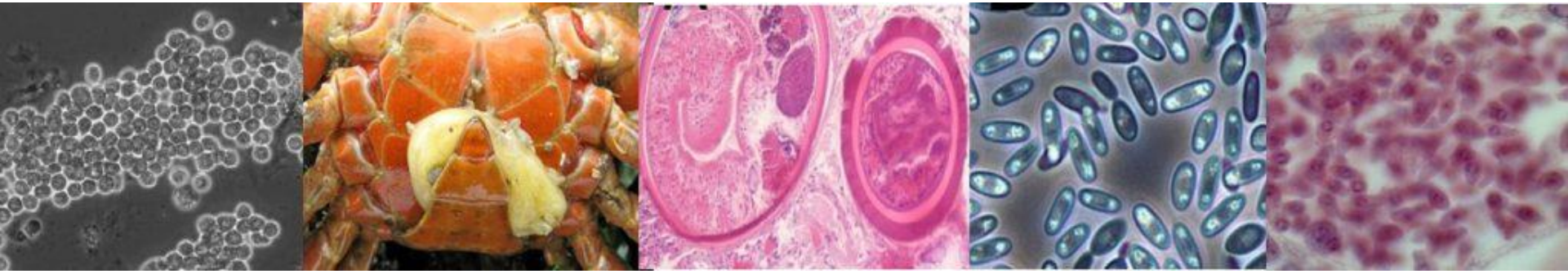
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Pathogen	<i>C. maenas</i>		Seawater eDNA	
	Dock	Pier	Dock	Pier
<i>Haplosporidia</i>	X	✓	✓	✓
<i>Microsporidia</i>	✓	X	✓	X
Mikrocytids	X	X	✓	✓
Paramyxids	X	X	X	✓
<i>Vibrio</i> spp.	✓	✓	✓	✓
Fungal species	X	✓	✓	✓
Trematode parasites	X	✓	NA	NA
<i>Sacculina</i> sp.	✓	X	NA	NA

Davies et al. (2021) Preprint. BioRxiv DOI: [10.1101/2021.05.26.445743](https://doi.org/10.1101/2021.05.26.445743)



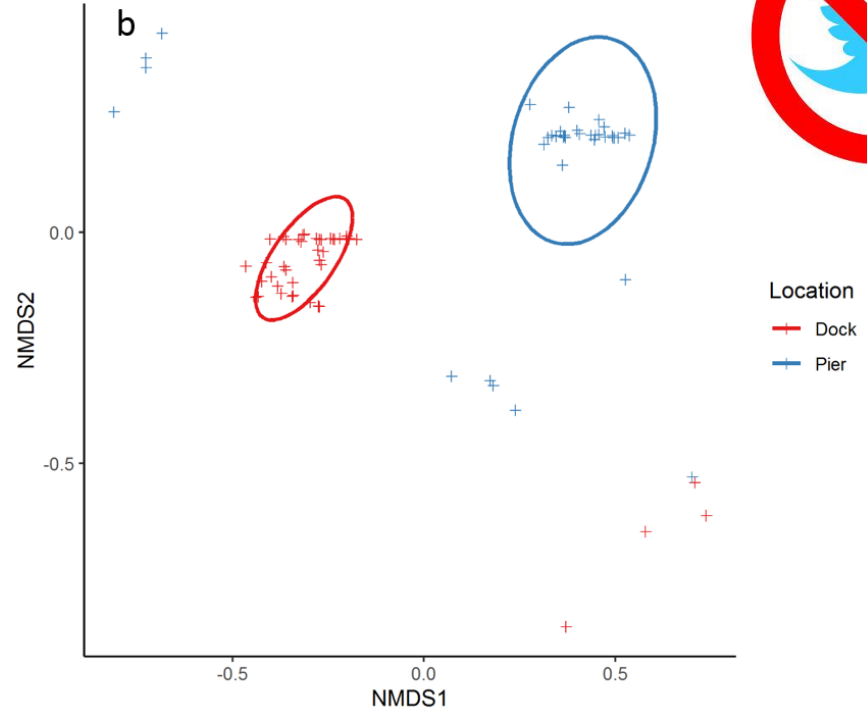
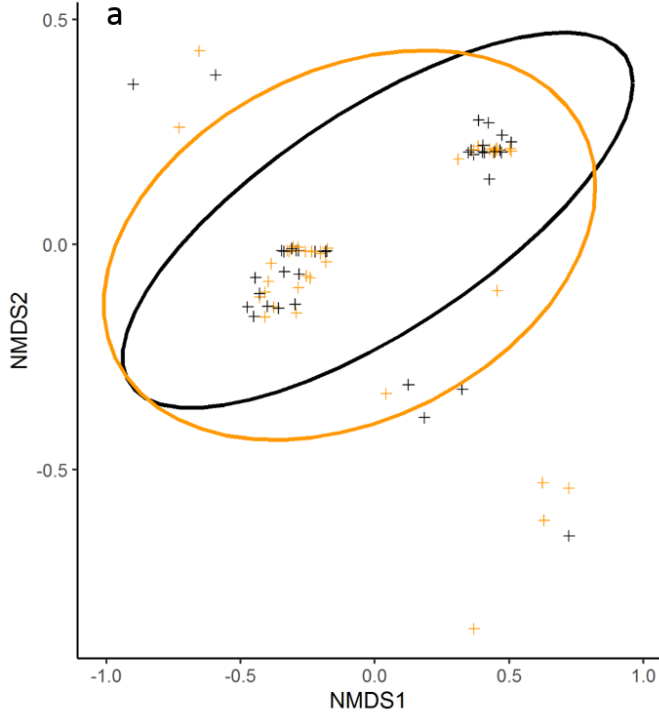
Co-infections



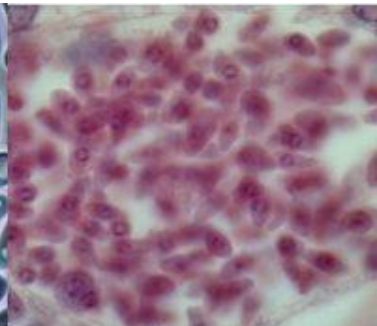
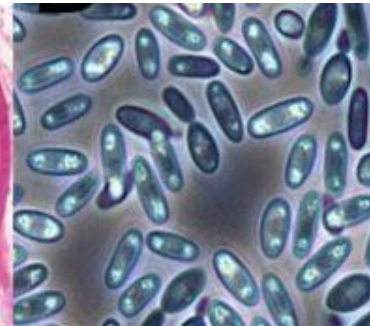
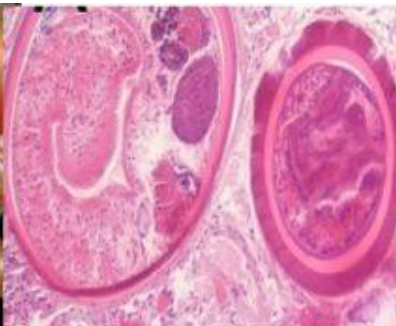
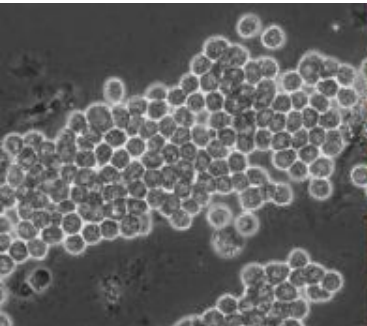
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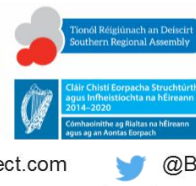
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Davies et al. (2021) Preprint. *BioRxiv* DOI: [10.1101/2021.05.26.445743](https://doi.org/10.1101/2021.05.26.445743)



Conclusions

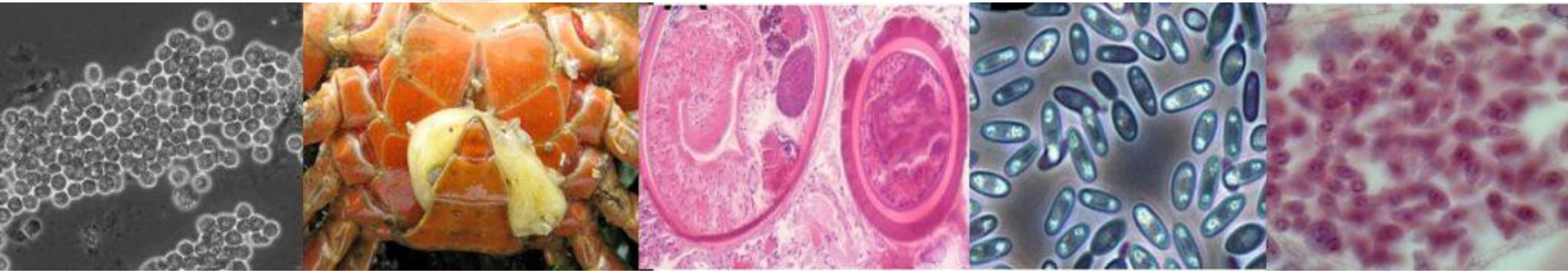


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- A systematic approach is always best
 - Using multiple detection methods
- eDNA can reveal difference stages of life cycles which may not be present in the host
- Location/environment may play a role in co-infection and disease dynamics
 - As opposed to only the host ecology



Thank you

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 @CharlotteEve

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- Frederico Batista, David Bass, Kelly Bateman and Stephen Feist at **Cefas**.
- Georgia Ward at the **Natural History Museum**.
- **Davies et al. (preprint)** Environment, rather than *Hematodinium* parasitization, determines collateral disease contraction in a crustacean host. *BioRxiv* DOI: [10.1101/2021.05.26.445743](https://doi.org/10.1101/2021.05.26.445743)
- **Davies et al. (2020)** Diagnosis and prevalence of two new species of haplosporidians infecting shore crabs *Carcinus maenas*: *Haplosporidium carcini* n. sp., and *H. cranc* n. sp. *Parasitology* **147**:1229–1237 DOI: [10.1017/S0031182020000980](https://doi.org/10.1017/S0031182020000980)
- **Davies et al. (2020)** Mycosis is a Disease State Encountered Rarely in Shore Crabs, *Carcinus maenas*. *MDPI Pathogens* **9**(6):462 DOI: [10.3390/pathogens9060462](https://doi.org/10.3390/pathogens9060462)
- **Davies et al. (2019)** Spatial and temporal disease dynamics of the parasite *Hematodinium* sp. in shore crabs, *Carcinus maenas*. *Parasites & Vectors* **12**:472 DOI: [0.1186/s13071-019-3727-x](https://doi.org/0.1186/s13071-019-3727-x)

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