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Training course 2014

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18th Annual Workshop of the National Reference Laboratories for Fish Diseases

DTU Vet National Veterinary Institute

2 training courses available for september 2014

The courses are entitled:

- Methods for implementation of surveillance procedures for listed fish diseases week 37 from Monday the 8th to Friday the 12th of September

- Real time PCR for diagnostics and surveillance of fish diseases in week 38 from Monday the 15th to Wednesday 17th of September

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...Learning from the experience...

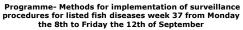
- Courses presentation

- Information on economy
- Registration procedure



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Section 1 FISH farm inspection	Section 2 Laboratory introduction	Section 3 Cell culture facility	Section 4 qPCR analysis	Section 5 Discussion and evaluation
FISH FARM VISIT in	9:00-10:00	8:30-10:00	8:30 - 12:00	9:00-10:00
collaboration with	Participants will be	Cell culture preparation for	Nucleic acid purification	Scientific discussion and
FVST (Danish	introduced to the	diagnostic purpose, titration	and master mix	recommendations. Final
Veterinary and Food	laboratory	and IFAT	preparation	results obtained, conclusion
Administration, Section				
for Aquaculture). The	Coffee Break 10:00-10:30	Coffee Break10:00-10:30		Coffee Break10-10:30
participants will be	10:30-12:00			
introduced to the	Theoretical introduction to	10:30-12:00 Titration		10:30-12:00
contingency plan in	the use of cell culture and	procedure		Course Evaluation and
Denmark, practical	qPCR for surveillance			feedback from participants
demonstration of farm	programs for non-exotic			
inspection will be	listed disease in Europe.			
performed and samples	Lunch 12 -12:30	Lunch12 -12:30	Lunch12 -12:30	Lunch12 -12:30
performed and samples for surveillance will be		Lunch12 -12:30 12:30- 14:30 Cell observation	Lunch12 -12:30 12:30-14:30	Lunch12 -12:30 12:30-15:00
performed and samples for surveillance will be collected by	Lunch 12 -12:30 12:30 - 14:30 Samples processing collected			
performed and samples for surveillance will be	Lunch 12 -12:30 12:30 - 14:30 Samples		12:30-14:30	12:30-15:00
performed and samples for surveillance will be collected by	Lunch 12 -12:30 12:30 - 14:30 Samples processing collected		12:30-14:30	12:30-15:00 Last minutes questions and
performed and samples for surveillance will be collected by	Lunck 12 -12:30 12:30 - 14:30 Samples processing collected following different		12:30-14:30	12:30-15:00 Last minutes questions and Good Byes Wrapping up of the course and questionnaire fill out
performed and samples for surveillance will be collected by	Lunch 12-12:30 12:30 – 14:30 Samples processing collected following different diagnostics procedure (Cell		12:30-14:30	12:30-15:00 Last minutes questions and Good Byes Wrapping up of the course
performed and samples for surveillance will be collected by	Lunch 12-12:30 12:30 – 14:30 Samples processing collected following different diagnostics procedure (Cell		12:30-14:30	12:30-15:00 Last minutes questions and Good Byes Wrapping up of the course and questionnaire fill out
performed and samples for surveillance will be collected by	Lunck 12–12–30 12:30 – 14:30 Samples processing collected following different diagnostics procedure (Cell culture and PCR)	12:30- 14:30 Cell observation <u>Coffee Break 14:30-15:00</u>	12:30-14:30 qPCR analysis <u>Coffee Break 14:30-15:00</u>	12:30-15:00 Last minutes questions and Good Byes Wrapping up of the course and questionnaire fill out
performed and samples for surveillance will be collected by	Lunck 12–12–30 12:30 – 14:30 Samples processing collected following different diagnostics procedure (Cell culture and PCR)	12:30- 14:30 Cell observation <u>Coffee Break 14:30-15:00</u> 15:00-16:30	12:30-14:30 qPCR analysis <u>Coffee Break 14:30-15:00</u> 15:00-16:30	12:30-15:00 Last minutes questions and Good Byes Wrapping up of the course and questionnaire fill out
performed and samples for surveillance will be collected by	Lunck 12–12–30 12:30 – 14:30 Samples processing collected following different diagnostics procedure (Cell culture and PCR)	12:30- 14:30 Cell observation <u>Coffee Break 14:30-15:00</u>	12:30-14:30 qPCR analysis <u>Coffee Break 14:30-15:00</u> 15:00-16:30 PCR results to be	12:30-15:00 Last minutes questions and Good Byes Wrapping up of the course and questionnaire fill out
performed and samples for surveillance will be collected by	Lunck 12–12–30 12:30 – 14:30 Samples processing collected following different diagnostics procedure (Cell culture and PCR)	12:30- 14:30 Cell observation <u>Coffee Break 14:30-15:00</u> 15:00-16:30	12:30-14:30 qPCR analysis <u>Coffee Break 14:30-15:00</u> 15:00-16:30	12:30-15:00 Last minutes questions and Good Byes Wrapping up of the course and questionnaire fill out

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Working in the lab cells and PCR



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Learning objectives

Maintain, cultivate, inoculate and read most used cell lines (BF-2, EPC, CCB and ASK) for fish disease diagnostic purposes.

Prepare cell cultures for different purposes, e.g. diagnosis, IFAT and virus titration.

Inoculate and sub-cultivate diagnostic samples.

Read diagnostic trays.

Titrate virus.

Apply Real-Time PCR for surveillance purposes.

Assess pitfalls and errors in test performances and designs.

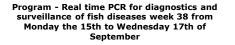
The major focus will be on the viral fish diseases using VHS as model.

The course will provide a forum where pre-knowledge, experience and examples can be discussed between participants and teachers, and hereby raise the awareness of pitfalls when using the various techniques.

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		Day 3
Section 1 Basic PCR	Section 2 primers and probes	Section 3
9:00-9:30 Registration and presentation of	9:00-10:00 Chemistry	9:00-9:45 Analysis and thresholds
9:30-10:00 Presentation of participants	(SYBR green, Taqman, FRET, others)	9:45-10:30 Cut-offs
10:00-10:45 History of PCR	10:00-10:30 coffee	10:30-11:00 coffee
10:45-11:15 Coffee	10:30-12:00 Controls and standards	A CONTRACTOR OF A CONTRACT OF A CONTRACT.
11:15-12:00 Practical recommendations	(melting curves, standard curves, internal controls, reference genes)	11:00-12:00 interpretation and presentation
(lab setup, instruments, plasticwares, handling of samples)	-	
12:00 - 13:00 Lunch	12:00 - 13:00 Lunch	12:00 - 13:00 Lunch
Section 1 Sample prep and validation	Section 2	Section 4
Section 1 Sample prep and validation 13:00-13:45 Sample preparation	Section 2 13:00-14:00 Primer/probe design in silico	Section 4 13:00-14:30 Theoretical exercises
13:00-13:45 Sample preparation		13:00-14:30 Theoretical exercises
13:00-13:45 Sample preparation	13:00-14:00 Primer/probe design in silico	13:00-14:30 Theoretical exercises Analysis of results 14:30-15:00 General discussion and cours
13:00-13:45 Sample preparation Extraction RNA/DNA 13:45-14:15 Preventing contamination (UNG)	13:00-14:00 Primer/probe design in silico	13:00-14:30 Theoretical exercises Analysis of results
13:00-13:45 Sample preparation Extraction RNA/DNA	13:00-14:00 Primer/probe design in silico 14:00-14:30 Coffee	13:00-14:30 Theoretical exercises Analysis of results 14:30-15:00 General discussion and cours evaluation
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Guest teachers

Marc Engelsma Dr.ir. CVI, Netherlands



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Learning objectives

The participants that will have fully followed all the course's objectives should be able to:

Validate new real-time PCR techniques for the lab.

Design and evaluate new primer/probe sets.

Understand the basic chemistry behind PCR.

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Understand how to analyse and interpret the results. Knowledge of the $\ensuremath{\mathsf{MIQE}}$ guidelines.

Troubleshooting.

The major focus will be on assays for listed fish diseases.

The course will provide a forum where pre-knowledge, experience and examples can be discussed between participants and teachers, and hereby raise the awareness of pitfalls when using the technique

Presentation name 17/04/2008

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Information on economy

This year the EU commission has allocated a specific budget for organizing and holding these training courses meaning that we can offer up to 15 participants from NRLs of EU Member States the following package: Package for up to 15 NRL representatives of EU Member states includes:

- No course fee

- Travel expenses up to 500 € will be refunded (economy fares)
- Up to 5 nights at hotel incl. breakfast will be paid and booked by the EURL
- Lunches and coffee breaks will be included

- 1 social dinner

For other members of governmental institutions such as Universities, Veterinary institutes etc. the package includes: - No course fee

- Travel expenses and accommodation must be covered by the participants themselves

- Fee for lunch and coffee of around 25 ${\ensuremath{\mathbb C}}$ per day plus fee for social dinner

Finally members of private companies are welcome to apply for participation. The package offered to these participants includes: - Course fee (700 C for course in week 37 and 600 C for course in week 38)

- Travel expenses, hotel and daily allowances to be covered by the participants themselves

- Fee for lunch and coffee of around 25 € per day plus fee for social dinner

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Application process roadmap

• From May 19th to July 1st REGISTRATION OPEN visit http://www.eurl-fish.eu/Activities/traning/Training_courses_2014



European Union Reference Laboratory for Fish Diseases

Registration Form for Training Courses 2014

And

motivational letter

to be delivered to <u>niven@vet.dtu.dk</u>

July 10th PARTICIPATION CONFIRMED TO APPLICANTS

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Thanks for your attention we look forward to welcome you here in September!

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