



Adaptation and Validation of the Purcell method of IHNv detection



L.Louboutin, J.Cabon, M.Baud, L.Bigarré, T. Morin

Efficiency

Between 91 and 96% (slopes ranging from -3,4 to -3,5)

Repeatability

	1 operator, 2 days						
	Nb of sessions		Operators		Nb of dilutions		Replicates/dilution
	2	1	6	2	6	2	2
Concentrations (PFU/ml)	1.10 ⁷	1.10 ⁶	1.10 ⁵	1.10 ⁴	1.10 ³	1.10 ²	1.10 ¹
06/05/2015	15,76	18,36	22,31	25,72	28,92	31,7	
12/05/2015	15,78	18,51	22,48	25,99	29,43	32,45	
Mean value	15,77	18,44	22,4	25,86	29,12	32,08	
Standard deviation	0,08	0,15	0,11	0,16	0,36	0,72	
CV%	0,51%	0,82%	0,51%	0,62%	1,22%	2,25%	

CV < 3%

3 operators

	Within operators						
	Nb of sessions		Operators		Nb of dilutions		Replicates/Dilution
	3	3	6	2	6	2	2
Concentrations (PFU/ml)	1.10 ⁷	1.10 ⁶	1.10 ⁵	1.10 ⁴	1.10 ³	1.10 ²	1.10 ¹
EG 07/05/2015	16,3	18,77	22,77	26,26	29,57	33,11	
LI 06/05/2015	17,35	19,96	23,74	27,3	29,85	32,2	
LM 06/05/2015	16,98	19,81	23,52	26,71	28,98	31,28	
Mean value	16,88	19,52	23,34	26,76	29,47	32,20	
Standard deviation	0,49	0,58	0,46	0,4899	0,40	0,93	
CV%	2,93%	2,96%	1,95%	1,83%	1,36%	2,89%	

CV < 6%

Perspectives

- 1 production of RNA Transcript to determine precisely LD_{PCR} (copies of RNA/reaction)
- 2 Add an exogenous control to validate the whole process (extraction + amplification)

Choice - bacteriophage T4 (DNA) and MS2 (RNA) to spike samples before extraction
 Diagnostic sensibility : 10 fold serial dilutions with and without control

- 3 Reproducibility : interested in participating a proficiency test for IHNv detection

- 4 To be accredited for IHNv detection by RT-PCR in september 2017
 Transfer the method to the French accredited labs (7 labs)

Then deployment of the range of analysis (VHSV and KHV in 2018 ? IPNV in 2019 ?) while maintaining accreditation in cellular virology ...

Protocol

Qiagen Quantitect Probe RT-PCR kit

Primer and probe concentrations → Purcell et al (900nM of each primers, 200nM of probe)

Reagents	Initial Concentration	Final Concentration	Volume for 1 reaction	Volume for 9 reactions	Programme
RT Probe master mix	2x	1x	19,50	176,50	1. 30 min 50°
pVP377	20	900	1,13	10,13	2. 15 min 95°
pVP378	20	900	1,13	10,13	3. 15 sec 94°
pVP23	20	200	0,250	2,25	4. 60 sec 60° ***
RT RT mix			0,250	2,25	FAM
H ₂ O			4,75	42,75	5. goto 3. 39x
Volume total (µl)			20,00	180,00	
Add	5	µl ABN to	20	µl MasterMix	

Tests of ruggedness

- Slight variations for :
 - Primers and probe concentrations (+/- 20%)
 - PCR program
- Two-step versus one-step
- 5 ten-fold dilutions (supernatant of N61, internal French reference strain)

No significant CT difference with +/- 20% [primers] or [probe]

Analytical specificity

40 supernatants

Status delivered by the method	Sample status	
	Positive	Negative
Positive	34 true positive	0 false positive
Negative	0 False Negative	6 True Negative

100% positive supernatant detected

Analytical sensitivity (limit of detection)

LOD	Nb of séances	Operators	Nb of dilutions	Replicates/dilution
DNA plasmid	3	1	6	8
N61 RNA	3	1	6	2

- 25 copies / reaction using plasmid pVPV22 (Ct=33,4)
- 10 PFU / reaction using N61 IHNv strain (2.10³ PFU/mL); Ct=32,6

Diagnostic specificity : Dsp = 100%

Status delivered by the method	Sample status	
	Positive	Negative
Positive	8 true positive	0 false positive
Negative	0 False Negative	2 True Negative

Diagnostic sensitivity

- 2 methods :
 - organ homogenate spiked with plasmidic DNA = 10 cp / reaction (Ct=34,5)
 - organs collected after experimental infections ; serial dilutions (Ct=35,2)
- Quite difficult to find Dse (random signal with late Ct)