

**3.** A primary cell line from allogynogenetic silver  
“crucian” carp, *Carrassius auratus gibelio* brain  
(**CrCB**) (Duan Hongan)  
and progress in CyHV-2 research

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May 30, 2017 Copenhagen

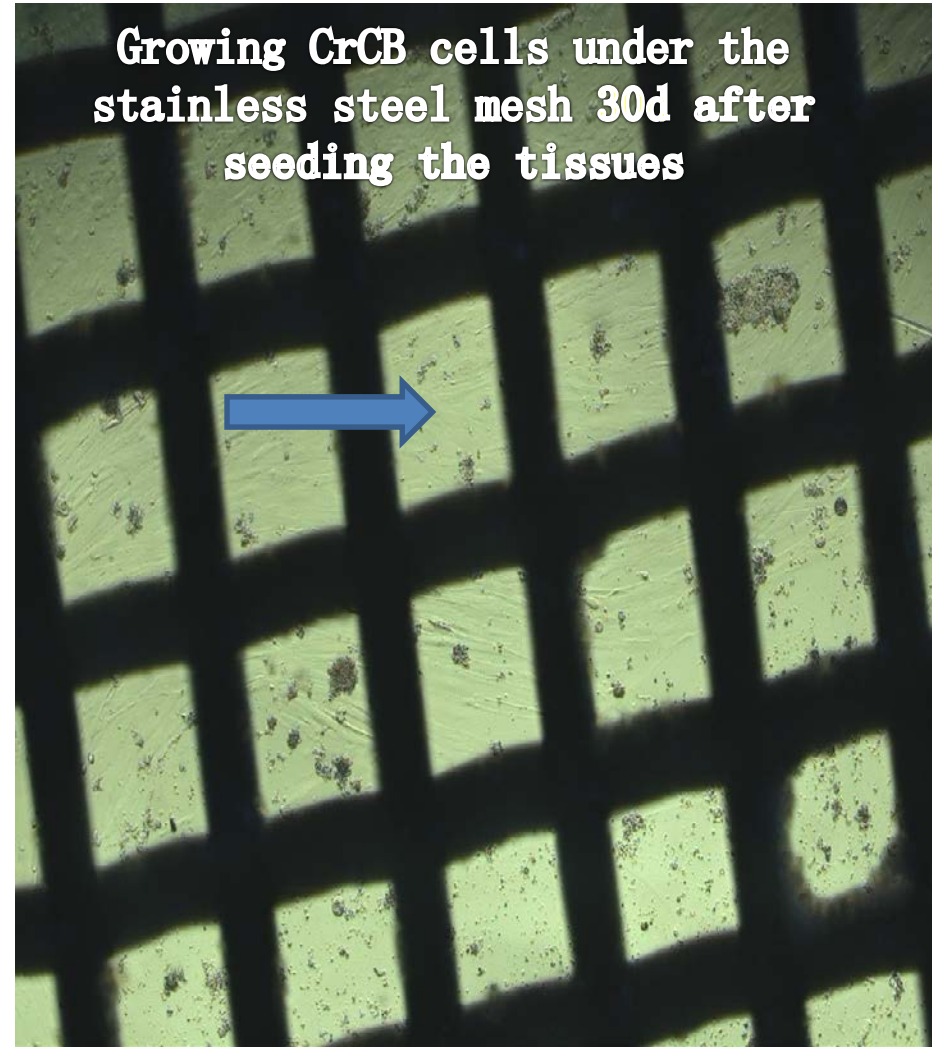
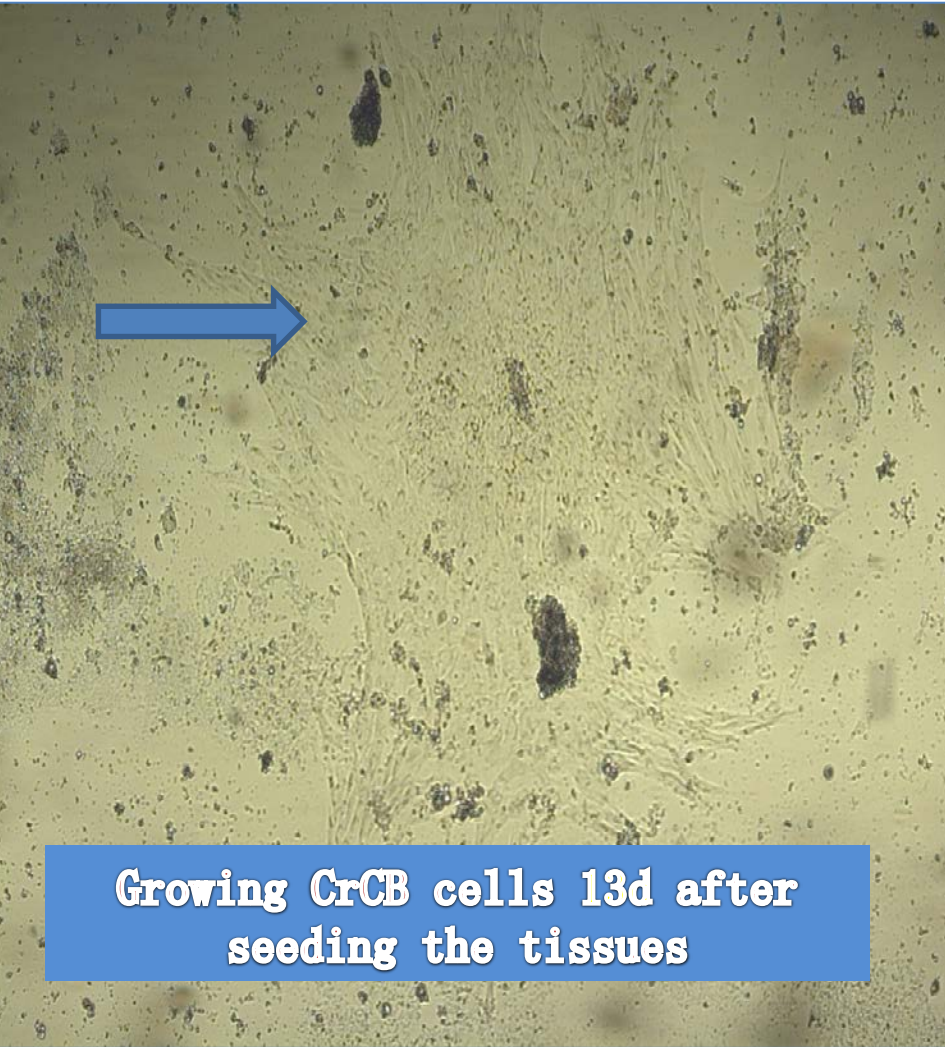
# Main contents

- Establishment of primary cell line from allogynogenetic *Car. auratus gibelio* brain
- Experimental infection of goldfish and PCR testing for CyHV-2
- Isolation of CyHV-2 strains from goldfish, and PCR
- Research in the future
- Acknowledgements

## SOP Establishment of primary cell line from allogynogenetic *Car. auratus gibelio* brain

- Aseptically take brain from healthy allogynogenetic silver crucian carp (*Carassius auratus gibelio*) of 8-10cm long, rinse 2x in 75% ethanol, 10-15 sec each, rinse 3x in sterile MEM or M199 medium
- Cut brain tissue into about 1mm<sup>3</sup>, transfer into 15ml sterile centrifuge tube, add 2 times the volume of trypsin solution, incubate about 5 min at 25 °C
- Centrifuge for 2 min at 2000 rpm, discard the supernatant
- Suspend the pellet in appropriate amount of MEM medium with 10% FBS, centrifuge for 2 min at 2000 rpm, discard the supernatant
- Transfer the pellet into the cell culture flask with a sterile inoculation ring or other equipment, plate the tissue to a single layer, cover the tissue block with a sterile stainless steel mesh with wire diameter 0.20 mm and aperture 0.308 mm, incubate flask at 25°C for 6-24h. Add M199 medium containing 20% FBS, penicillin 100 U/ml, streptomycin 0.1mg/ml, FGF (fibroblast growth factor) 10µg/ml and/or EGF (epidermal growth factor) 10µg/ml. Replace the fresh medium every 3-5 days
- When ~80% confluence: subculture the cells, then passage every 10 days
- From passage # 10 replace medium with 10% FBS, but no antibiotics, FGF and EGF.
- Passage the cells ~every 5 days.

# Primary cell line from *C.aur.gibelio* brain



## Further tests on these cells

- 16S and 18S goldfish mtDNA fragment primers were used to differentiate the cell lines. The amplified products were sequenced and analysed by Blast analysis.
- Giemsa staining, and the cell chromosome number and karyotype analysis were done.

# Establishment of primary cell line from allogynogenetic *Car. auratus gibelio* brain

## Results

- The first generations of CrCB cells adhered difficult to the flask and grew slowly into a confluent monolayer
- It took ~ 30 days for the CrCB cells to grow to ~80% confluence, from then cell growth gradually accelerated. From the 10<sup>th</sup> passage it took 24-48h for the cells to grow into confluent monolayers
- CrCB has a fibrous cell morphology
- PCR showed target fragments (123bp and 234bp) that were amplified by 16S and 18S primers and confirmed by sequencing and blast analysis
- Chromosome analysis and further characterization is in progress

# Experimental infection with and isolation of CyHV-2 from goldfish

- Feb 2017 goldfish kept at 3-6°C were taken to the lab and put in the tank, and the water was heated to about 25 °C
- Brain kidney and spleen (**BKS**) and intestine of sick and dead fish ➡ PCR test
- If PCR + for CyHV-2 ➡ homogenized and filtered with 0.45µm filter and inoculated onto the CrCB
- CPE positive CrCB tested by PCR for CyHV-2 and sequencing



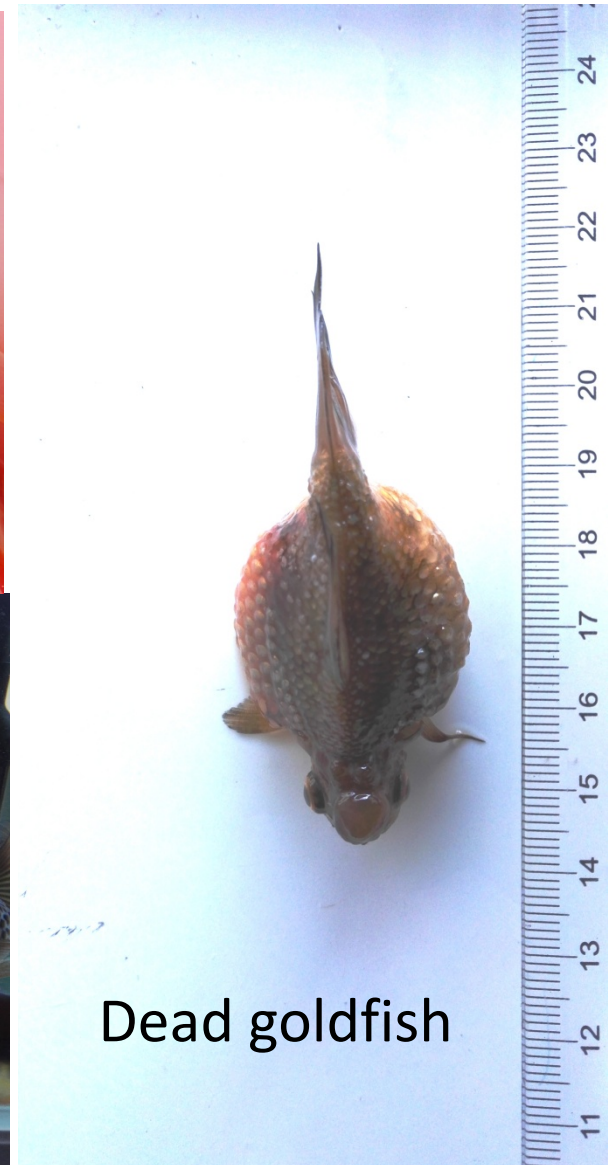
# Experimental infection of and Isolation of CyHV-2 from goldfish



PCR- goldfish



PCR+ goldfish

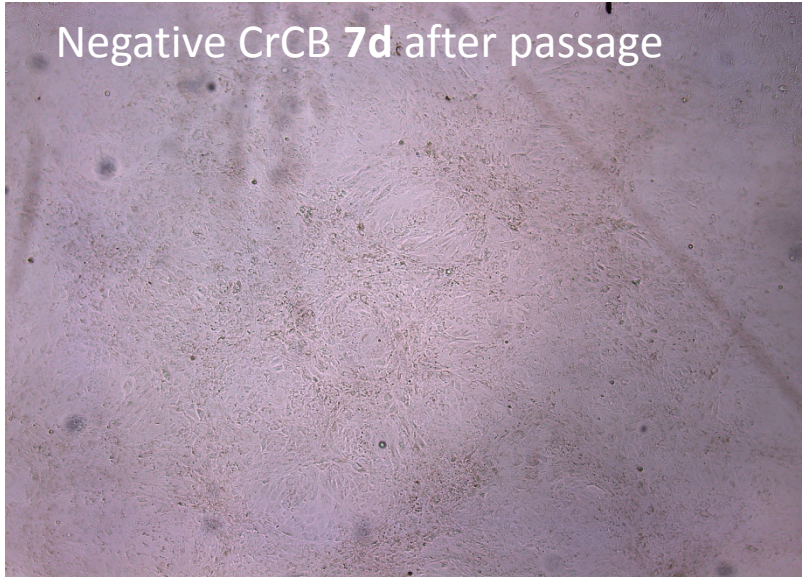


Dead goldfish

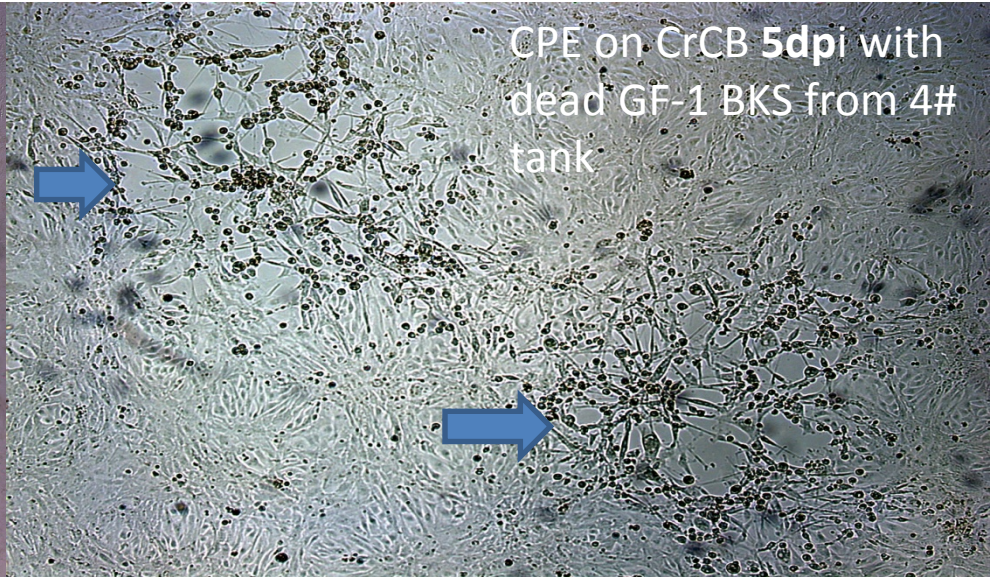


# Experimental infection of and Isolation of CyHV-2 from goldfish

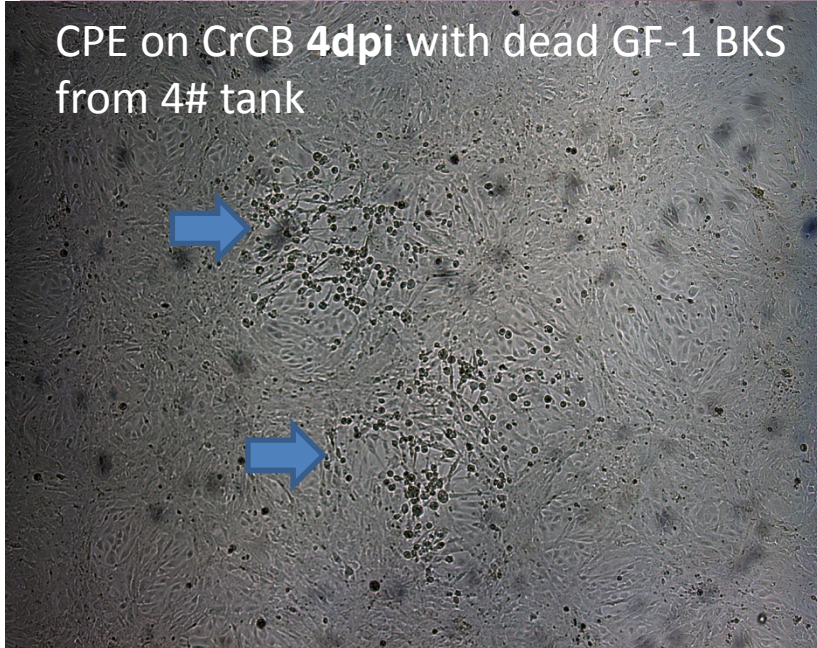
Negative CrCB 7d after passage



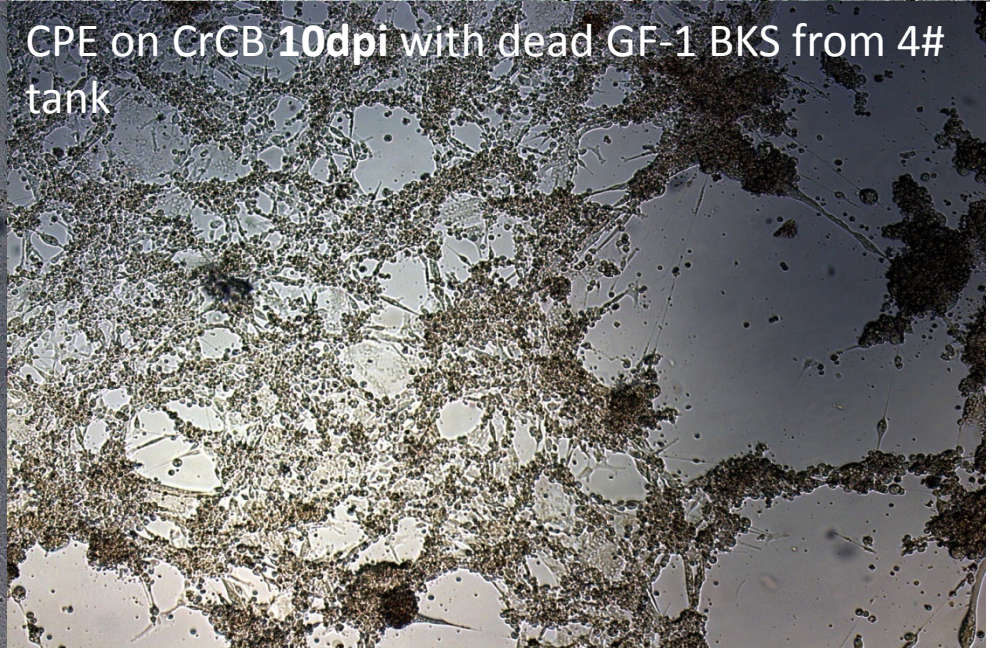
CPE on CrCB 5dpi with dead GF-1 BKS from 4# tank



CPE on CrCB 4dpi with dead GF-1 BKS from 4# tank



CPE on CrCB 10dpi with dead GF-1 BKS from 4# tank





# Experimental infection of and Isolation of CyHV-2 from goldfish

## Results

- Fish began to die from day 6 p.i. (PCR+ for CyHV-2)
- Tissues from sick or dead goldfish: PCR + for CyHV-2
- CrCB cells inoculated with tissue homogenates from sick/dead goldfish showed CPE at 3-5d post-inoculation.
- CPE+ CrCB suspensions were PCR positive for CyHV-2

# Research in the future

- Further characterization of CrCB
- Further characterization of CyHV-2 strains
- Production of reagents for detection of CyHV-2
- Production of inactivated CyHV-2 vaccine and trial

# Acknowledgements

- Thank you for your attention!
- Thanks for the organizers to give the opportunity for the inclusion the presentation in this conference
- Thanks for Dr Olga Haenen to edit and present this PPT to the meeting
- Welcome to research cooperation on diagnosis and prevention of this disease
- CrCB and CyHV-2 strains can be sent to you on request

# Contact

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0086-13951258955

# Characterization of primary cell line CrCB

reference: Establishment, characterization, and viral susceptibility of two cell lines derived from goldfish *Carassius auratus* muscle and swim bladder (Rougée et al., 2007)

16S fragment (123 bp) :

16S-F 5'-GCGACCACGGAGGAAAAA-3'

16S-R 5'-CGTTGATCGGCTTGTATTAG-3'



M: DL2000; 1: CrCB; 2: goldfish brain cell; 3-5: negative control

**Fig1. 16S PCR results**

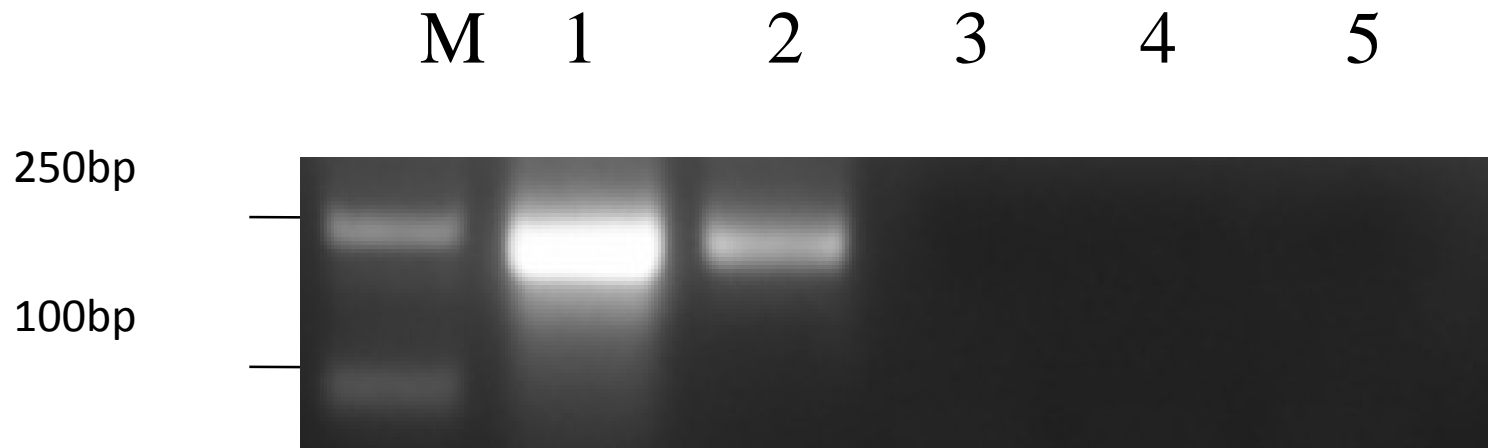


# Characterization of primary cell line CrCB

reference: Establishment, characterization, and viral susceptibility of two cell lines derived from goldfish *Carassius auratus* muscle and swim bladder (Dis Aquat Org 77: 127–135, 2007)

18S fragment (234 bp) :

(18S-F)5'-GCGAGACGAGCCACCACCTATC-3' (18S-R) 5'-CCCCCGGCCGTCCCTCTTA-3'.



M: DL2000; 1: CrCB; 2: goldfish brain cell; 3-5: negative control

fig2. **18S PCR results**

# Characterization of primary cell line CrCB

## Sequences producing significant alignments:

Select: [All](#) [None](#) Selected:0

[Alignments](#) [Download](#) [GenBank](#) [Graphics](#) [Distance tree of results](#)

Description	Max score	Total score	Query cover	E value	Ident	Accession
<input type="checkbox"/> <a href="#">Carassius gibelio isolate 2 mitochondrion, complete genome</a>	159	159	100%	6e-36	100%	<a href="#">KU896992.1</a>
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<input type="checkbox"/> <a href="#">Carassius gibelio isolate CQBP2 mitochondrion, complete genome</a>	159	159	100%	6e-36	100%	<a href="#">KU668573.1</a>
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## 16s F alignment result

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Select: [All](#) [None](#) Selected:0

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<input type="checkbox"/> <a href="#">Carassius gibelio isolate 1 mitochondrion, complete genome</a>	158	158	98%	2e-35	100%	<a href="#">KU896991.1</a>
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<input type="checkbox"/> <a href="#">Carassius gibelio isolate CQBA1 mitochondrion, complete genome</a>	158	158	98%	2e-35	100%	<a href="#">KU668568.1</a>
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## 16s R alignment result

# Characterization of primary cell line CrCB

## Sequences producing significant alignments:

Select: [All](#) [None](#) Selected:0

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<input type="checkbox"/> <a href="#">Carassius gibelio 18S ribosomal RNA gene, partial sequence</a>	285	285	99%	3e-73	94%	<a href="#">KT002365.1</a>
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<input type="checkbox"/> <a href="#">Carassius auratus 18S ribosomal RNA gene, partial sequence</a>	285	285	99%	3e-73	94%	<a href="#">FJ710819.1</a>
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## 18s F alignment result

## Sequences producing significant alignments:

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[Alignments](#) [Download](#) [GenBank](#) [Graphics](#) [Distance tree of results](#)

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<input type="checkbox"/> <a href="#">Carassius auratus 18S ribosomal RNA gene, partial sequence</a>	241	241	98%	6e-60	90%	<a href="#">FJ710819.1</a>
<input type="checkbox"/> <a href="#">Carassius auratus gibelio 18S ribosomal RNA gene, partial sequence</a>	241	241	98%	6e-60	90%	<a href="#">EF189737.1</a>
<input type="checkbox"/> <a href="#">Cyprinus carpio 18S ribosomal RNA gene, partial sequence</a>	235	235	98%	3e-58	89%	<a href="#">KT002364.1</a>
<input type="checkbox"/> <a href="#">Cyprinus carpio genome assembly common carp genome, scaffold 000028946</a>	235	706	98%	3e-58	89%	<a href="#">LN591197.1</a>
<input type="checkbox"/> <a href="#">Onychostoma macrolepis 18S ribosomal RNA gene, partial sequence</a>	235	235	98%	3e-58	89%	<a href="#">JQ283288.1</a>
<input type="checkbox"/> <a href="#">Cyprinus carpio intergenic spacer, partial sequence; 18S ribosomal RNA gene, internal transcribed spacer 1, 5.8S ribosomal RNA gene, internal transcribed spacer</a>	235	235	98%	3e-58	89%	<a href="#">JN628435.1</a>
<input type="checkbox"/> <a href="#">Cyprinus carpio 18S ribosomal RNA gene, partial sequence</a>	235	235	98%	3e-58	89%	<a href="#">FJ710826.1</a>
<input type="checkbox"/> <a href="#">Cyprinus carpio intergenic spacer region, partial sequence; 5' external transcribed spacer, 18S ribosomal RNA gene, internal transcribed spacer 1, 5.8S ribosomal</a>	235	235	98%	3e-58	89%	<a href="#">AF133089.2</a>

## 18s R alignment result

# Experiment infection of and Isolation of CyHV-2 from goldfish

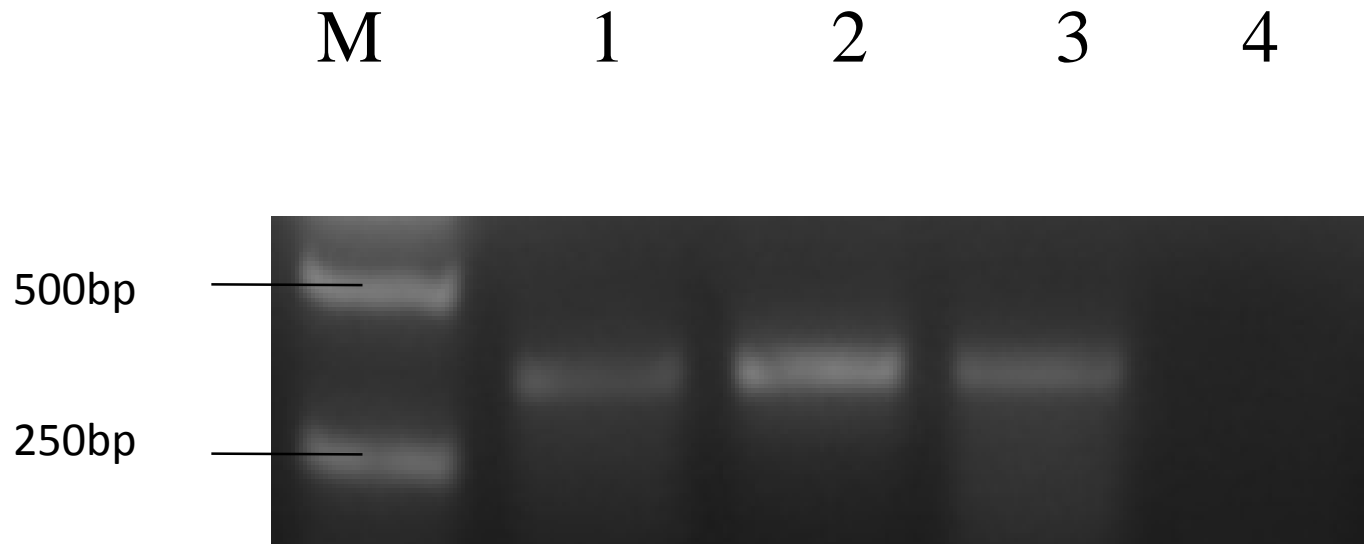
3# tank of gold fish			4# tank of gold fish			Tank of PCR- goldfish		
Total 13			Total 12			Total 13(divided after Feb24)		
Date	Death	PCR	date	death	PCR	4 in 3# tank	4 in 4# tank	5 control
Feb15	2	+	Feb18	1	+	1 death on	no death	no death
Feb 17	1 morbid	+	Feb21	1	+	Feb2 PCR-		
Feb18	1	+	Feb22	2	+	1 death on		
Mar8	1	+	Feb25	1	+	Mar14 PCR-		
Mar24	1	+	Feb26	1	+	1 death on		
			Mar13	1	+	May4 PCR-		
total	7		total	7				

## CyHV-2 PCR

Primers: CyHV pol-FOR ( CCCAG CAACATGTGCGACGG )

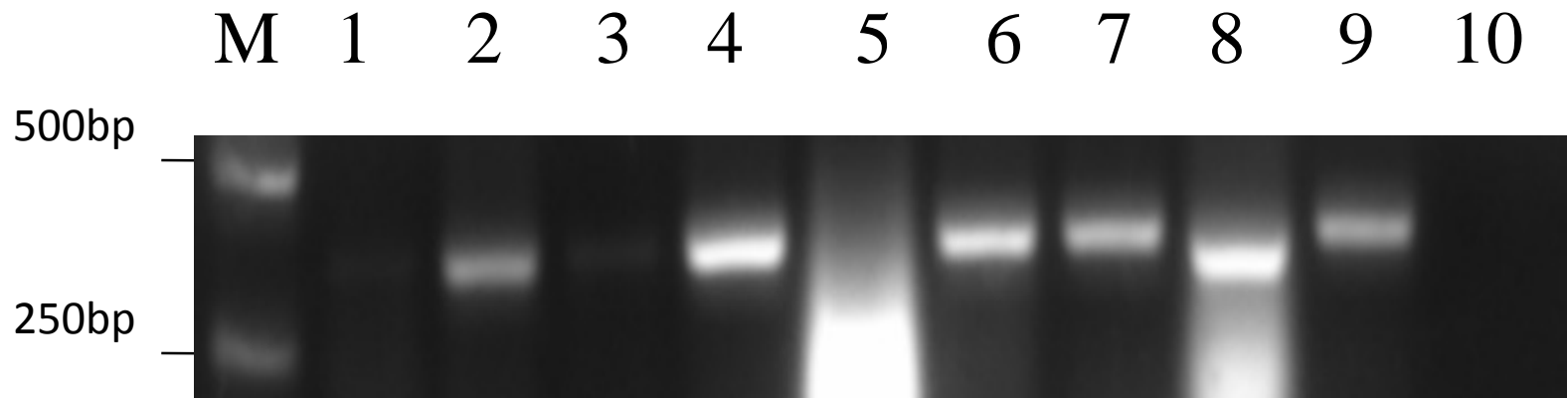
CyHVpol-REV ( CCGTARTGAGAGTTGGCGCA )

Fragment size: 362 bp



M: DL2000 ladder; lane 1: intestine; lane 2: brain kidney and spleen mixture ; lane 3: positive control; lane 4 blank control  
fig3. PCR results for dead goldfish on Feb 15,2017

# PCR results



M: DL2000 marker ; lane1:zzxz21(XUZHOU GF); lane2-6: BKS, intestine of GF died on Feb17 in 3# tank; intestine,BKS,brain of GF died on Feb18 in 3# tank; Lane7-8: intestine ,BKS of GF died on Feb18 in 4# tank; lane 9: positive control(PC); lane10: blank control(BC);

Note: GF-goldfish, DGF-dead goldfish, BKS –brain kidney and spleen  
Fig4. PCR results from cell suspension inoculated with infected tissue homogenate



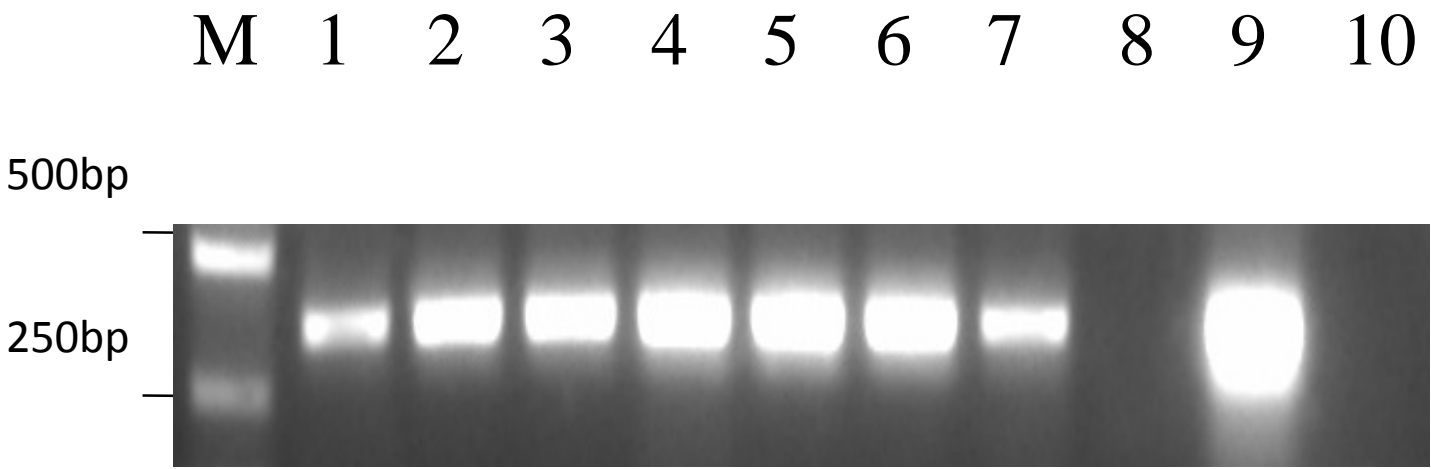
## PCR results for 2 dead goldfish from 4<sup>#</sup>tank



Lane 1,2: intestine, brain kidney and spleen from dead goldfish No.1;  
lane3,4: intestine, brain kidney and spleen from dead goldfish No.2;  
lane5:positive control; lane6 : blank control; M: DL2000

Fig6. PCR results for 2 dead goldfish from 4<sup>#</sup>tank

# 20170303 PCR results for CrCB suspension infected by BKS of DGF of DGF



M: DL2000; lane1-8: CrCB suspension infected by BKS of DGF from 4# tank died and inoculated on the CrCB on Feb21(1,2) ,Feb22 (3, 4) , Feb25 (5, 6) ,Feb26 (only brain, 7, 8) ; lane9: positive control; lane10: Blank control; BKS: brain kidney and spleen mix; DGF: dead goldfish

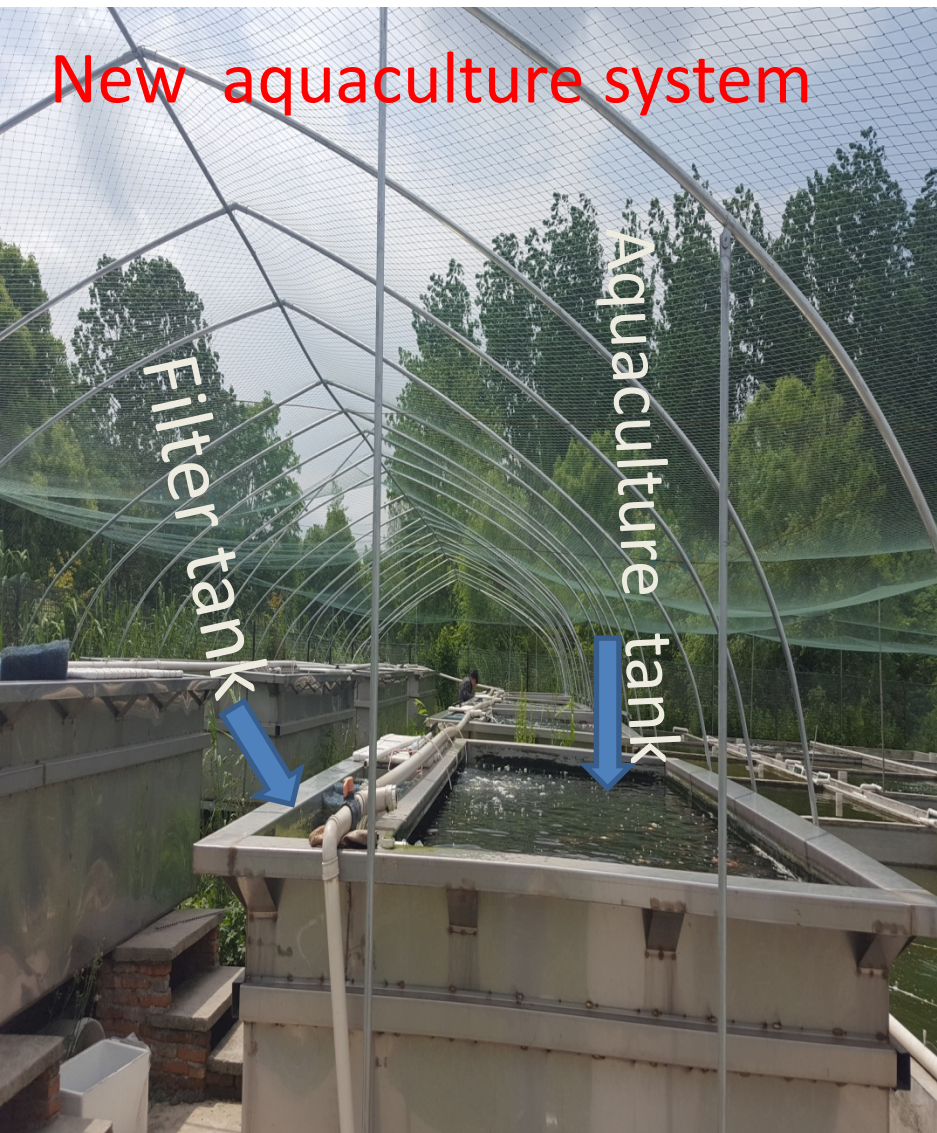
Fig7. PCR results for CrCB suspension infected by BKS of DGF from 4# tank

## Example farm for prevention trial of CyHV-2

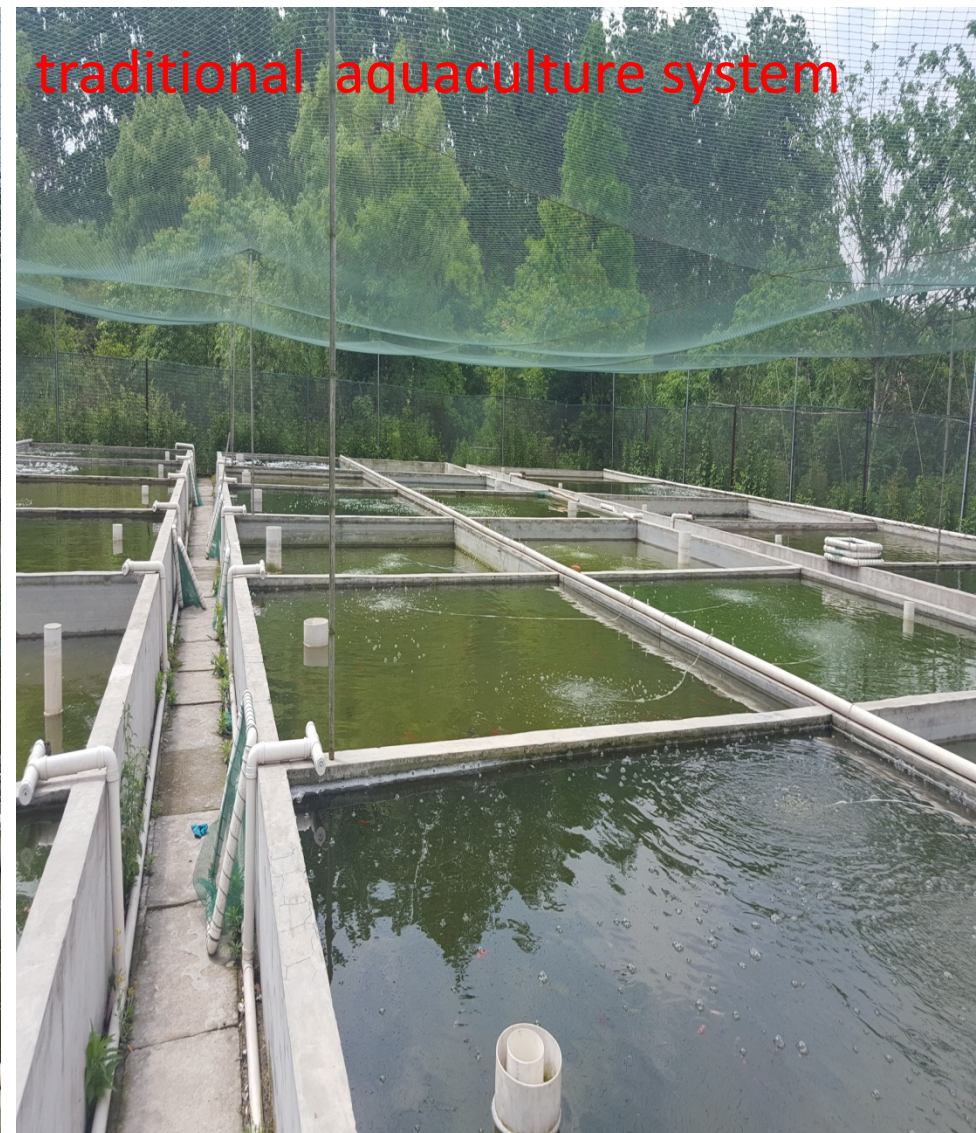
- Suzhou AquaBridge Aquarium Technology Co., Ltd is a registered company by AQSIQ for export of ornamental fish mainly goldfish.
- The company is under the control of local CIQ. The target surveillance and pre-export test is done by this laboratory.
- The company has developed a new system and good operation procedure for prevention of CyHV-2 and other diseases.

# example farm for prevention trial of CyHV-2

New aquaculture system



traditional aquaculture system





# example farm for prevention trial of CyHV-2



- Filter tank and aquaculture tank are integrated
- Water through 3 layers of filter is pumped into the aquaculture tank
- Green *Wolffia arrhiza* serves as plant feed source and to reduce organic ingredients( faeces, feed residue et al)
- **one species one pool from the fry stage**
- avoidance of introduction of diseases by self-breeding
- strict control policies and sound hygiene practices
- Isolated quarantine area for export of ornamental fish

