



IHN outbreak in rainbow trout farmed in Saltwater in Croatia

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FARMING OF RAINBOW TROUT IN SALTWATER

NASLOVNA OBALA OTOCI TURIZAM NAUTIKA RIBOLOV KOLUMNE MAGAZIN INFORMACIJE

Kreće projekt uzgoja pastrve u Velebitskom kanalu

Prije 2 godine



Naslovnica / Zadar / Podvelebitški ribari u moru love – pastrve

Zadar Zadarskožupanija

Podvelebitški ribari u moru love – pastrve

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POČETNA DOGAĐAJI KRONIKA ZADAR ŽUPANIJA KULTURA SPORT ZABAVA PRILOZI MAG

Početak - Svjetska promocija uzgoja pastrve u Jadranu

Svjetska promocija uzgoja pastrve u Jadranu

0 Srt, 10/08/2011 - 00:00 - 1 - Podijeli: Facebook Twitter

Autori: Damir Marčić



Brama pirata u moru kao i okus nepopovijni su i nadmoći rječnom uzgoju

Djelomično u sjeni lijepog i značajnog čina - promocije vina Zadarske županije na Donat wine festivalu, a koju su nam upriličili iz Udruge vinogradara i vinara Zadarske županije "Vina Liburna", objavljena je još jedna ujedno i svjetska promocija proizvoda ovog kraja - pastrva uzgojena u Podvelebitskom kanalu.



- several attempts to establish farming of rainbow trout in the Adriatic Sea, mostly unsuccessful
- during the summer temperatures are up to 25°C causing low oxygen saturation
- Velebitki Kanal was evaluated as one of several suitable sites for this project
- area with an abundant inflow of submerged rivers supplying marine environment with cold freshwater making this site suitable for rainbow trout farming



Lesson learnt from past:

- rainbow trout should be placed into cages during the favourable weather during winter time (no wind, low sea temperature)
- very good growth rate
- Desirable flavour of the market size fish
- Depletion of oxygen during the summer months – trigger for different bacterial diseases

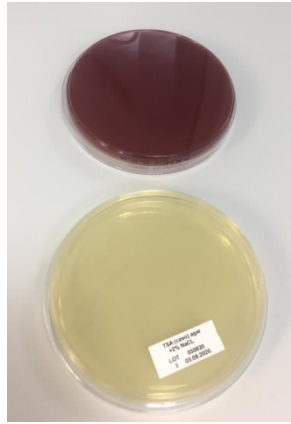
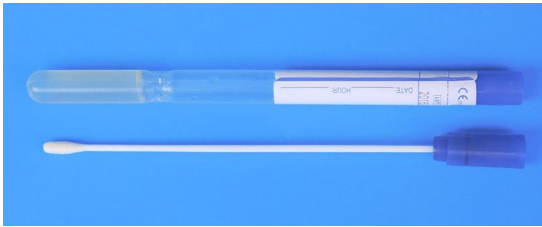
POLAND

DENMARK



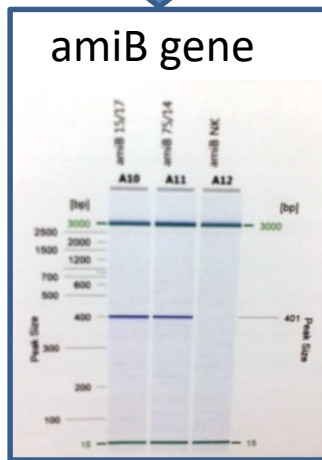
DIAGNOSTICS

Bacteriology



Vibrio spp.

Bacterial DNA was isolated from pure culture using NucleSpin Microbial DNA kit (Macherey Nagel, Germany) according to the manufacturer instructions



Vibrio anguillarum



Virology

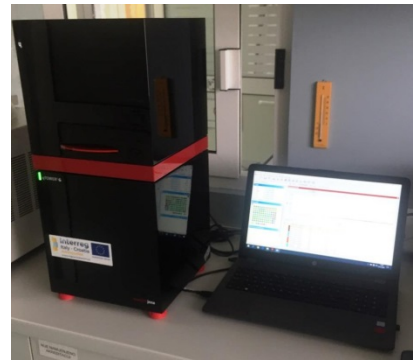


Viral RNA was extracted on KingFisher Duo Prime Purification System (Thermo Scientific) using MagMAX CORE Nucleic Acid Purification Kit following the manufacturers simple workflow instructions for processing animal swab samples

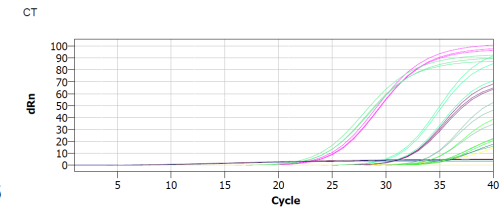
RT-q-PCR for detection of VHSV according to Commission decision 2015/1554



VHSV negative



Modified Purcell et al. (2013) single step RT-q-PCR for detection of IHN



C11	PK IHN 18/3 p.t	Positive control	FAM		24.53	24.35
A10	57/20-5 bris	Unknown	FAM		31.62	30.9
A11	57/20-5 bris	Unknown	FAM		30.68	30.9
A12	57/20-5 bris	Unknown	FAM		30.42	30.9
E7	57/20-1 bris	Unknown	FAM		31.9	31.91
E8	57/20-1 bris	Unknown	FAM		32.01	31.91
E9	57/20-1 bris	Unknown	FAM		31.83	31.91
F7	57/20-2 bris	Unknown	FAM		23.57	23.43
F8	57/20-2 bris	Unknown	FAM		23.64	23.43
F9	57/20-2 bris	Unknown	FAM		23.09	23.43
G7	57/20-3 bris	Unknown	FAM		34.03	34.34
G8	57/20-3 bris	Unknown	FAM		34.9	34.34
G9	57/20-3 bris	Unknown	FAM		34.09	34.34

Notification of IHNV presence in saltwater environment to CA

- Official visit to the farm and sampling of fish in different cages; C1(10), C2(7), C3(8),C4(5)
- Specimens submitted to the laboratory weighed 50-300 grams



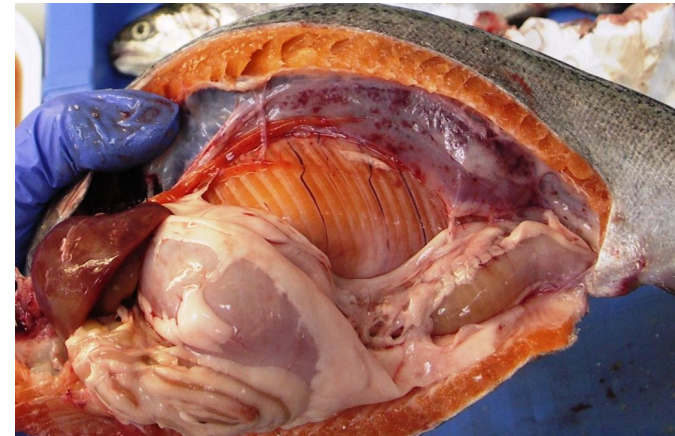
Hemorrhages on the skin of the opercula, vent, fin bases



In the mouth



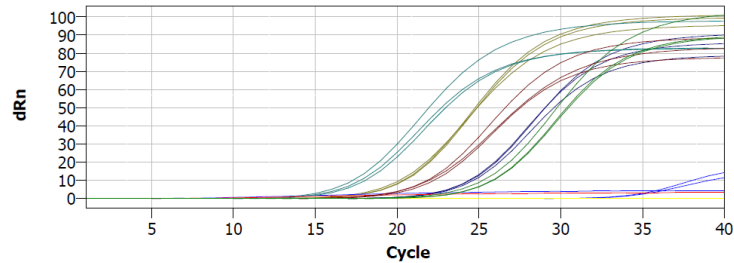
Dark gills and liver, hemorrhages in pyloric caeca, liver, enlarged spleen, swim bladder



DIAGNOSTIC PROCEDURES:

- Fish from each cage were considered as a pool
- Material from organs was inoculated on EPC and BF 2 cell lines and CPE appeared 6th day after inoculation for sample C1, & C4 on EPC and sample C3 on both EPC and BF2 while sample C2 was negative even after subcultivation in new cell culture
- Positive supernatants were tested for the presence of VHSV, IHNV and IPNV using commercial ELISA kits (TestLine, Czech Republik; BioX, Belgium)
- Samples C1 & C4 tested positive for IHNV while C3 tested positive for IPNV
- ELISA IHNV positive samples were tested using RT-qPCR (single step modified protocol by Purcell et al. 2013)
- ELISA IPNV positive sample was tested using end-point PCR targeting VP2/NS junction region (Heppell et al. 1992)

CT

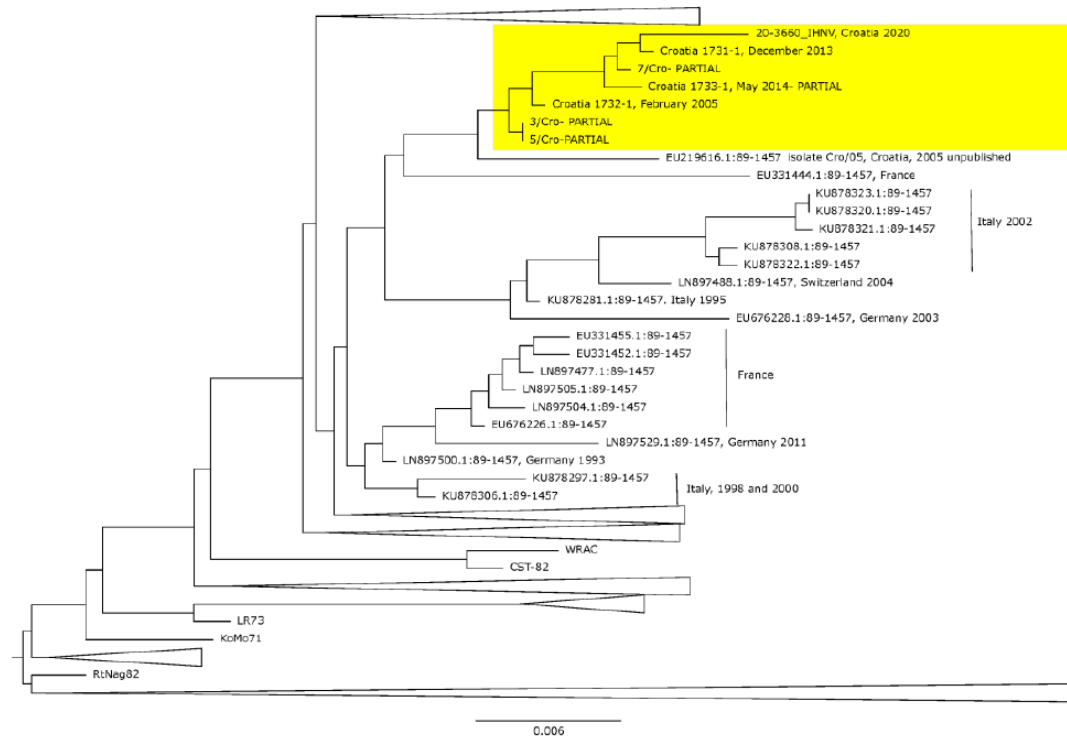


Well		Sample name	Sample type	Dye	Gene	Ct	Mean Ct
A1		61/20-1 E	Unknown	FAM		19.03	19.22
A2		61/20-1 E	Unknown	FAM		19.27	19.22
A3		61/20-1 E	Unknown	FAM		19.34	19.22
B1		61/20-3 E	Unknown	FAM		No Ct	
B2		61/20-3 E	Unknown	FAM		36.38	36.65
B3		61/20-3 E	Unknown	FAM		36.92	36.65
C1		61/20-4 E	Unknown	FAM		23.29	23.39
C2		61/20-4 E	Unknown	FAM		23.53	23.39
C3		61/20-4 E	Unknown	FAM		23.34	23.39
D1		61/20-1 B	Unknown	FAM		16.17	16.6
D2		61/20-1 B	Unknown	FAM		16.63	16.6
D3		61/20-1 B	Unknown	FAM		16.99	16.6
E1		61/20-3 B	Unknown	FAM		No Ct	
E2		61/20-3 B	Unknown	FAM		No Ct	
E3		61/20-3 B	Unknown	FAM		No Ct	
F1		61/20-4 B	Unknown	FAM		20.7	20.83
F2		61/20-4 B	Unknown	FAM		21.11	20.83
F3		61/20-4 B	Unknown	FAM		20.7	20.83
G1		NTC	NTC	FAM		No Ct	
G2		NTC	NTC	FAM		No Ct	
G3		NTC	NTC	FAM		No Ct	
H1		PK	Positive control	FAM		24.11	24.52
H2		PK	Positive control	FAM		24.71	24.52
H3		PK	Positive control	FAM		24.73	24.52

- amplification and sequencing of „mid-G“ region of the G gene of the IHN virus (Kolodziejek et al. 2008) confirmed the identification and showed similarity to CRO/05 in Genbank

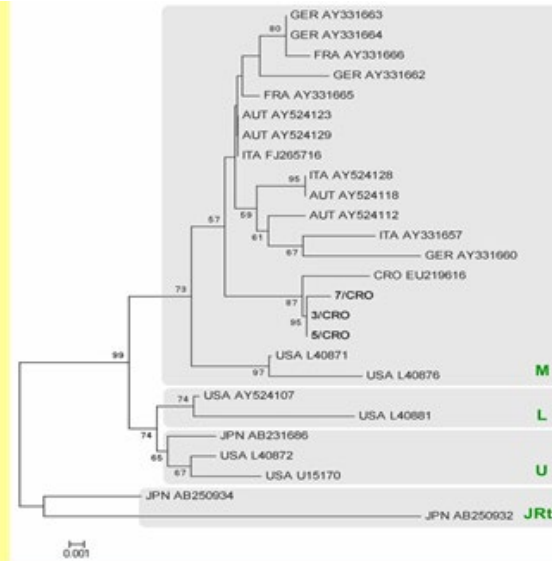
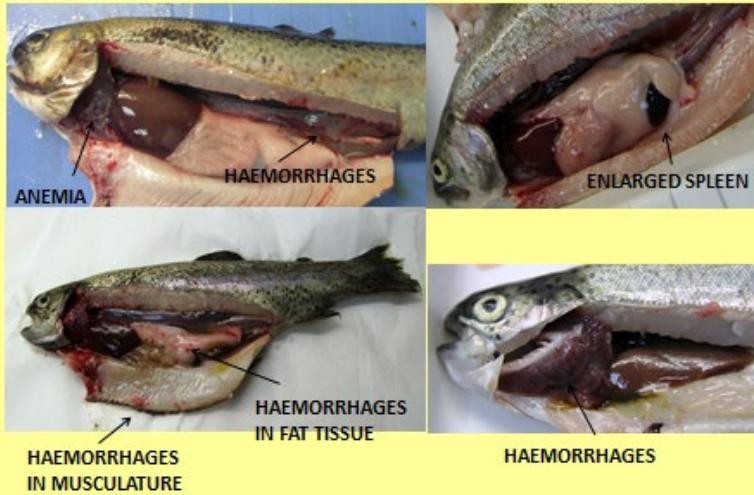
Results of sequencing and phylogeny

- Different samples (organ suspension, supernatant from EPC cell lines) were sent to EURL for confirmation – all sequences were the same – IHNV genotype E

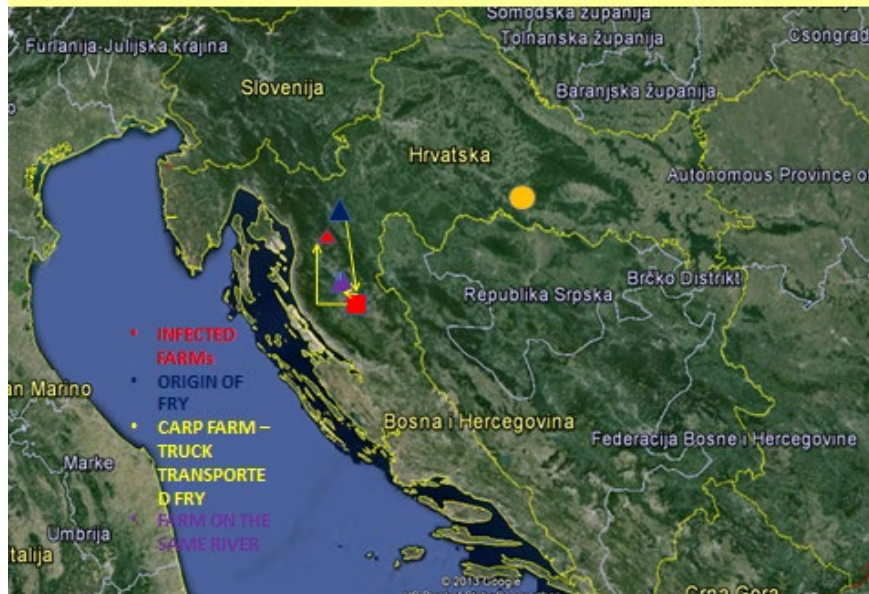


- Comparison of sequences available in Genbank database and sequences from unpublished Croatian outbreaks in 2005, 2013 & 2014 confirmed this isolate is the closest to CRO/05 but also closely correlates with other national isolates

Findings

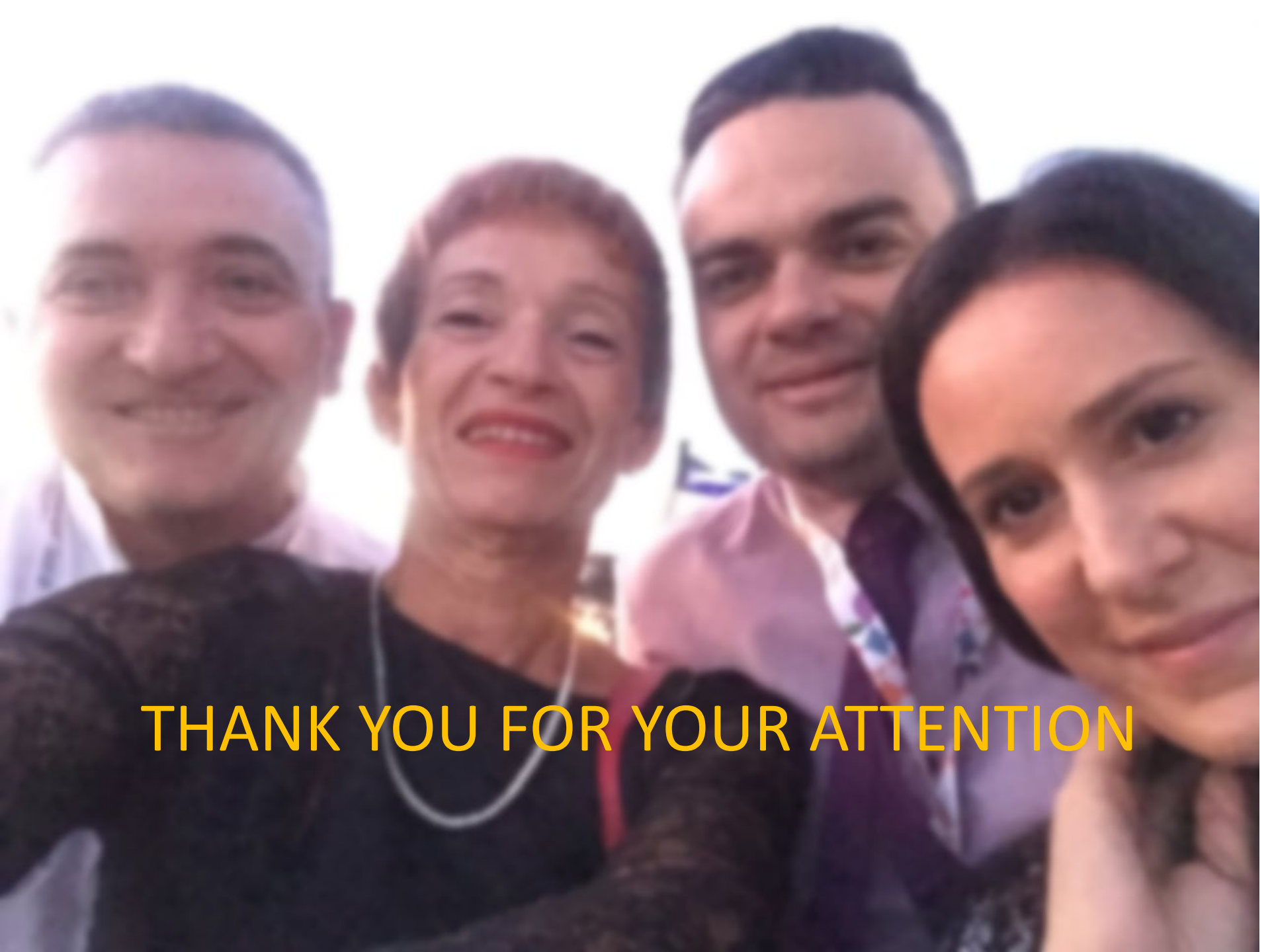


MOVEMENT OF FISH AND VEHICLE



Conclusion:

Based on the data obtained by partial sequencing of glycoprotein gene are suggesting the possibility of circulation of the virus within the country rather than being imported from other countries



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